Pd(II)-Catalyzed, Picolinamide-Assisted, Z‑Selective γ‑Arylation of Allylamines To Construct Z‑Cinnamylamines

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ABSTRACT: Investigations of Pd(II)-catalyzed, picolinamide-assisted, γ-C(sp²)−H activation and Z-selective arylation of allylamines are reported. The reactions of N-allylpicolinamides with various aryl iodides in the presence of the catalyst $Pd(OAc)₂$ and additive AgOAc have led to the selective γ arylation of allylamines to construct various cinnamylamines with moderate to good yields and good to high E/Z ratios. To obtain good E/Z ratios, the Pd(II)-catalyzed arylation reaction

of N-allylpicolinamides was probed using different additives, directing groups, and reaction conditions. The Pd(II)-catalyzed arylation of an allylamine containing both γ -C(sp²)−H and γ -C(sp³)−H bonds afforded moderate yields of the γ -C(sp²)−H and γ-C(sp³)−H bisarylated cinnamylamines. Although Heck-type γ-arylations of allylamines have generally afforded the Ecinnamylamines, the bidentate directing group picolinamide-directed arylations of allylamines were found to be Z-selective. A plausible mechanism was proposed for the observed regioselectivity and Z-selective arylation of N-allylpicolinamides. Additionally, the Pd(II)-catalyzed arylation of an N-allyl-5-methylisoxazole-3-carboxamide afforded the E-cinnamylamines plausibly via a ligand-free Heck-type reaction mechanism.

■ INTRODUCTION

Over the past few decades, organic synthesis has experienced the advantages of the celebrated transition-metal-catalyzed C− C bond-forming cross-coupling reactions (e.g., Kumada, Negishi, Stille, Suzuki, Hiyama, and Heck reactions) of suitable coupling partners.^{[1](#page-14-0)−[3](#page-14-0)} In recent years, the construction of C−C bonds via transition-metal-catalyzed C−H bond functionaliza-tion has received special attention^{[4](#page-14-0)-[8](#page-14-0)} because the C−H functionalization process offers additional advantages. For example, transition-metal-catalyzed C−H bond functionalization is a direct method for forming C−C bonds and does not require the preassembly of organometallic reagents. Often, C− H functionalization can be performed using commercially available starting materials (e.g., arenes, alkenes, cycloalkanes, carbonyls, and amines). The functionalization of sp^2 and sp^3 C−H bonds of organic molecules has been performed with or without the help of directing groups using transition-metal-based catalysts (e.g., Pd, Ru, Rh, Cu, and Ni).^{4-[10](#page-15-0)} Among the available C−H functionalization reactions, directing-group assisted C−H functionalization reactions have received substantial attention. $9,10$ $9,10$ $9,10$ This is because high levels of regioselectivity as well as stereoselectivity can be achieved.^{[9](#page-14-0)[,10](#page-15-0)} C−H functionalization of sp² or sp³ C−H bonds of organic molecules using Daugulis's bidentate directing group^{[11](#page-15-0)} (e.g., DG-a and DG-b), Yu's monodentate directing group,^{[12](#page-15-0)} and other related/modified bidentate directing groups have been extensively studied [\(Scheme 1\)](#page-1-0).^{[9](#page-14-0),[10,13](#page-15-0)−[21](#page-15-0)}

Allylamine derivatives are an important class of compounds, are ubiquitous in biologically active molecules, and are

considered important in medicinal chemistry research and pharmaceuticals.[22,23](#page-15-0) Allylamine derivatives or allylaminetethered molecules are notably useful synthetic building blocks for assembling various nitrogen-containing compounds.^{[24](#page-15-0)-[26](#page-16-0)} Various γ-arylated allylamine derivatives (cinnamylamines) have been found to exhibit a wide range of biological activities and have been identified as potential drug candidates. Notably, some cinnamylamine molecules are currently being used as medicines ([Figure 1\)](#page-1-0).^{[22,23](#page-15-0)} For example, naftifine is an antifungal drug used for the topical treatment of tinea pedis, tinea cruris, and tinea corporis. Flunarizine is a calcium antagonist, and flunarizine is effective in the prophylaxis of migraine. Cinnarizine has been characterized as an antihistamine and calcium-channel blocker and is prescribed for nausea and vomiting. Additionally, the cinnamylamine derivative CP-724,714 was found to stop the growth of tumor cells.

In general, allylamine derivatives are synthesized using various methods, and given the importance of cinnamylamines (γ-arylated allylamine derivatives) in medicinal chemistry research, considerable efforts have been made to assemble cinnamylamines.[27](#page-16-0)−[30](#page-16-0) Notably, the celebrated Mizoroki−Hecktype reaction $31-33$ $31-33$ $31-33$ comprising allylamines and suitable coupling partners (e.g., aryl halides or pseudohalides) is well utilized to assemble cinnamylamines.[34](#page-16-0)−[43](#page-16-0) Cinnamylamines have also been prepared via the metal-catalyzed C−H functionalization of allylamines by using arenes^{$42,43$ $42,43$ $42,43$} and the oxidative Heck-type

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Scheme 1. Bidentate Directing Group Assisted C−H Functionalization of Carboxylic Acids and Amines

Figure 1. Biologically active allylamines/cinnamylamines (γ-arylated allylamine derivatives).

reaction comprising allylamines and aryl boronic acids.^{[39,40](#page-16-0)} Typically, in these reactions, the corresponding cinnamylamines with E-geometries are obtained as predominant isomers. Consequently, the construction of Z-cinnamylamines as the predominant isomers has been less frequently encountered.^{[43](#page-16-0)} Furthermore, in some of the reactions, the Mizoroki−Hecktype arylation of allylamines was found to afford both the γ- and β -arylated allylamines.

A survey of the literature revealed that noteworthy investigations have been performed to accomplish regioselectivity in the Mizoroki−Heck-type arylation of allylamines. Most of these reactions afforded E-cinnamylamines as the major isomers.[34](#page-16-0)−[43](#page-16-0) Apart from the report by the group of Xu and Deng, 43 the construction of Z-cinnamylamines via the reaction of aryl halides and allylamines involving the transition-metalcatalyzed direct arylation technique (e.g., Heck and C−H activation reactions) 3^{34-43} 3^{34-43} 3^{34-43} 3^{34-43} 3^{34-43} has not been well explored. Recently, Liu reported^{[44a](#page-16-0)} the stereoselective synthesis of Z-allylic amines via a Cp_2TiCl_2 -catalyzed cis-hydroalumination of propargylic amines with Red-Al. Consequently, there exist only limited reports dealing with the construction of Z-cinnamylamines.

Our group recently reported $45a$ the stereoselective construction of Z-cinnamamide scaffolds via a $Pd(OAc)_{2}$ -catalyzed, bidentate directing group 8-aminoquinoline-assisted Z-selective β -C(sp²)−H arylation of acrylamides (eq 1, [Scheme 2](#page-2-0)). Taking

an impetus from this reaction and the existing developments regarding bidentate directing group assisted C−H functionalization, $9,10,13-18$ $9,10,13-18$ $9,10,13-18$ $9,10,13-18$ $9,10,13-18$ we envisaged the construction of Z-cinnamylamines via a Pd(II)-catalyzed, bidentate directing group and chelation-assisted Z-selective C−H activation followed by a γ- $C(sp^2)$ -H arylation of allylamines. To date, only two examples of bidentate directing group oxalylamide- and chelation-assisted Z-selective C−H functionalization of allylamines have been reported. $^{18c, d}$ $^{18c, d}$ $^{18c, d}$ The group of Yao and Zhao reported 18c an example comprising the synthesis of pyrrolidones via the γ- $C(sp²)$ -H carbonylation of allylamines (eq 2, [Scheme 2](#page-2-0)). The group of Wen and Zhao reported^{[18d](#page-15-0)} an example comprising the $\bar\gamma\text{-C}(\bar\text{sp}^2)\text{-H}$ silylation of allylamines (eq 2, [Scheme 2\)](#page-2-0). Herein, we report our investigations on the $Pd(OAc)₂/AgOAc$ catalytic system, picolinamide- and chelation-assisted Z-selective γ-C(sp²)-H arylation of N-allylpicolinamides (eq 3, [Scheme](#page-2-0) [2](#page-2-0)). This work divulges a contemporary method for the regioselective γ-arylation of allylamines involving relatively simple reaction conditions in which aryl iodide is a coupling partner, $Pd(OAc)_2$ is the catalyst, and AgOAc acts as an additive to regenerate the catalyst.^{[9](#page-14-0)[,10](#page-15-0),[13](#page-15-0)-[18](#page-15-0)}

■ RESULTS AND DISCUSSION

To begin our investigation on the Z-selective γ -C(sp²)-H arylation of allylamines, we assembled various N-allylcarboxour previous work:

bidentate directing group-aided β -arylation of acrylamide

acrylamide

available two examples:

bidentate directing group-aided γ -C-H functionalization of allylamine

amides to use as substrates in the $Pd(OAc)₂/AgOAc$, bidentate directing group assisted γ -C(sp²)−H arylation reactions. Accordingly, the N-allylcarboxamide substrates 1a−m were prepared from the corresponding allylamines and carboxylic acids (Scheme 3). After preparing the required N-allylcarboxamides 1a−m, we attempted the construction of Z-cinnamylamine 3a via the $Pd(II)$ -catalyzed arylation of N-allylpicolinamide 1a. [Table 1](#page-3-0) shows the optimization of the reaction conditions of the reaction of N-allylpicolinamide 1a with aryl halides 2a−c in the presence of various palladium catalysts, additives, and solvents.

The C−H arylation reaction of a mixture of Nallylpicolinamide 1a (1 equiv), 1-iodo-4-methoxybenzene (2a, 4 equiv), $Pd(OAc)_{2}$ (5 mol %), and AgOAc (2.2 equiv) in toluene at 110 °C afforded the γ -C(sp²)–H-arylated allylamine derivatives $3a'/3a$ (E/Z isomers) with a yield of 51% and an $E/$ Z ratio of 21:79 (entry 1, [Table 1](#page-3-0)). As was envisioned, this

reaction afforded the γ-C−H arylated allylamine derivative 3a with Z-stereochemistry as the major isomer, and this result indicated the involvement of a chelation-assisted mechanism of N-allylpicolinamide 1a. We next performed the same reaction using 10 mol % of the Pd(OAc)₂ catalyst, which afforded $3a'/$ 3a (E/Z isomers) with an improved yield (64%) and E/Z ratio of 11:89 (entry 2, [Table 1](#page-3-0)).

We then determined whether the yield and E/Z ratio of $3a'/$ 3a could be further improved using various palladium catalysts, additives, and solvents. The Pd(II)-catalyzed γ -C(sp²)–H arylation of 1a with 2a in the presence of Ag_2CO_3 afforded 3a′/3a with a yield of only 36% and an E/Z ratio of 8:92 (entry 3, [Table 1\)](#page-3-0). The arylation of 1a with 2a in the presence of PhI(OAc)₂ or KOAc failed to afford $3a'/3a$ (entries 4 and 5, [Table 1](#page-3-0)). The arylation of 1a with 2a in the presence of K_2CO_3 afforded $3a'/3a$ with a yield of only 29% and an E/Z ratio of 70:30 (entry 6, [Table 1](#page-3-0)). Notably, this reaction afforded the γ -C−H-arylated allylamine derivative 3a′ with E-stereochemistry as the major isomer, indicating the involvement of a conventional Heck-type reaction mechanism.^{[31,32,45a](#page-16-0)} We next attempted the arylation of $1a$ with $2a$ using $PdCl₂$ and $Pd(CH_3CN)_2Cl_2$ as the catalysts instead of $Pd(OAc)_2$. The reaction of 1a with 2a in the presence of $PdCl₂$ or $Pd(CH₃CN)₂Cl₂$ (10 mol %) and AgOAc (2.2 equiv) afforded 3a′/3a with yields of only 31−37% and an E/Z ratio of 11:89 (entries 7 and 8, [Table 1\)](#page-3-0). We also performed the arylation of 1a with 2a in different solvents, e.g., 1,2-dichloroethane (1,2- DCE), tert-butyl alcohol (t-BuOH), and tert-amyl alcohol (t-AmylOH). The Pd(II)-catalyzed reactions of 1a with 2a in 1,2- DCE and t-BuOH solvents were not fruitful (entries 9 and 10, [Table 1](#page-3-0)). The $Pd(II)$ -catalyzed reaction of 1a with 2a in t-AmylOH afforded 3a′/3a with a yield of only 37% and an E/Z ratio of 12:88 (entry 11, [Table 1\)](#page-3-0).

To improve the yield and E/Z ratio of $3a'/3a$, we screened the Pd(II)-catalyzed arylation of 1a using different amounts of 2a. Accordingly, the arylation of 1a (1 equiv) with 6 equiv of 2a afforded 3a′/3a in a marginally improved yield (69%) with an E/Z ratio of 4:96 (entry 12, [Table 1](#page-3-0)). The arylation of 1a (1) equiv) with 2−3 equiv of 2a afforded low yields of 3a′/3a (25− 48%) with slightly decreased E/Z ratios (E/Z ratio up to 16:84, entries 13 and 14, [Table 1](#page-3-0)). The arylation of 1a with the coupling partners 1-bromo-4-methoxybenzene (2b) and 1 chloro-4-methoxybenzene $(2c)$ instead of $2a$ did not afford

Scheme 3. Directing Groups and Substrates Employed for Investigating the γ-C(sp²)−H Arylation^{a,44b}

Table 1. Optimization of the Reaction of Allylamine 1a

Pdl ~ (5-10 mol %) additive (0.55 mmol) solvent (3 mL), 36 h. 80-110 °C $3a'$ (*E*-isomer) 3a (Z-isomer)

^aThe E/Z ratios were determined from the NMR spectra of the corresponding crude reaction mixtures. All of the reactions were performed using PdL₂ (5–10 mol %), additive (0.55 mmol), and solvent (3 mL) for 36 h unless otherwise stated. ^b 5 mol % of Pd(OAc)₂ was used. ^c6 equiv (1.5 mmol) of 2a was used. d_3 equiv (0.75 mmol) of 2a was used. e_2 equiv (0.5 mmol) of 2a was used. f The reaction was performed using 2b instead of 2a. ^gThe reaction was performed using 2c instead of 2a. h 0.27 mmol of 2a; the reaction time was 5 h. ⁱThe reaction time was 5 h. ^j0.83 mmol of 2a; the reaction time was 24 h. k_1 mmol of $2a$; the reaction time was 24 h.

3a′/3a (entries 15 and 16, Table 1). We also performed the arylation of 1a using various standard Heck-type reaction conditions, and these reactions were not fruitful (entries 17− 20, Table 1). The optimization reactions revealed that the isomers $3a'/3a$ were formed with different E/Z ratios under the experimental conditions. While we cannot ignore the occurrence of thermal cis−trans (Z/E) isomerization as discussed in our previous work, $45a$ a control reaction was performed to check whether the *cis–trans* (Z/E) isomerization is occurring under the thermal conditions or the E-isomer is formed via a conventional Heck-type reaction pathway. Accordingly, the isomers $3a'/3a$ ($E/Z = 1:99$) were treated with the catalyst $Pd(OAc)_{2}$ and additive AgOAc in the absence of 2a. The crude NMR of this reaction revealed a minor change in the E/Z ratio, and the E/Z ratio was found to be 11:89 (eq 1,

Table 1). Additionally, to check whether a second arylation of 3a′/3a occurs under the experimental conditions, we performed the arylation of $3a'/3a$ $(E/Z = 11:89)$ with iodobenzene. This reaction afforded a complex mixture, and purification of the crude mixture did not give the expected bisarylated product 3aA (eq 2, Table 1). After successfully achieving the arylation of substrates 1a, we then performed the Pd(II)-catalyzed arylations of substrates 1h−j, which were lacking the corresponding bidentate directing groups, and these reactions did not give the corresponding arylated products in characterizable amounts ([Scheme 3\)](#page-2-0).^{44b} These experiments indicated that, in substrate 1a, the corresponding bidentate directing group played an important role in the $Pd(\Pi)$ catalyzed C−H arylation process to afford 3a (major isomer).

Scheme 4. Pd(II)-Catalyzed Picolinamide- and Chelation-Assisted Construction of Z-Cinnamylamines 3b−m^a

 a The E/Z ratios were determined from the NMR spectra of the corresponding crude reaction mixtures, and in most cases, the corresponding major isomer (Z) was isolated in its pure form. ^bThese reactions were performed using the conditions of entry 12 in [Table 1](#page-3-0), and some other trials using the conditions of entry 12 in [Table 1](#page-3-0) were not fruitful. ²2 mmol of ArI was used. d In this case, compounds $3g'$ (*E*-isomer) and $3g$ (*Z*-isomer) were isolated in their pure forms. ^e2 mmol of ArI and 15 mol % of Pd(OAc)₂ were used. f_5 mol % of Pd(OAc)₂ was used.

With the reaction conditions optimized, we next wished to explore the generality and substrate scope of this protocol. Accordingly, Scheme 4 shows the arylation of 1a with a wide range of aryl iodides under the optimized reaction conditions (entry 2, [Table 1\)](#page-3-0). The arylation of 1a with aryl iodides containing an alkyl substituent at the para position afforded the corresponding γ-C−H arylated allylamines 3b−g with yields of 50−82% and moderate to good E/Z ratios (E/Z up to 9:91). The arylation of 1a with various disubstituted aryl iodides and an aryl iodide containing a substituent at the meta position (e.g., COOEt) afforded the corresponding allylamines 3h−k with yields of 40−59% and low to moderate E/Z ratios (E/Z) up to 21:79, Scheme 4). We also performed the arylation of 1a using another optimized reaction conditions (entry 12, [Table](#page-3-0) [1](#page-3-0)) in which 6 equiv of ArI (2a) was used. Accordingly, the products 3b, 3c, 3g, and 3h were obtained from the arylation of 1a with the corresponding aryl iodides with relatively good E/Z ratios (E/Z up to 15:85, Scheme 4).

We also performed the arylation of 1a using heteroaryl iodides to afford the corresponding allylamine derivatives 3l and 3m with yields of 50−69% and high E/Z ratios (E/Z up to 2:98, Scheme 4). Most of the allylamines 3′/3 were obtained in satisfactory yields (>50% yields) and E/Z ratios (E/Z ratios ranging from $38:62$ to 2:98). After performing the Pd(II)catalyzed γ-C−H arylation of allylamine 1a using picolinamide as the bidentate directing group to determine other working directing groups and obtain an improved yield and E/Z ratio, we attempted the γ-C−H arylation of allylamine 1b by using 4 chloropicolinamide as the bidentate directing group. The arylation of 1b with PhI, aryl iodides containing a substituent at the para or meta position (e.g., OMe, Me, Et, n-pentyl, nhexyl, i-Pr, and COOEt), disubstituted aryl iodides, and heteroaryl iodides afforded the corresponding γ-C−H arylated allylamine derivatives 4a−n with yields of 41−63% and low to good E/Z ratios (E/Z up to 12:88, [Scheme 5](#page-5-0)). Most of the derivatives 4′/4 were obtained with satisfactory yields (>50% yields) and E/Z ratios (E/Z) ratios ranging from 34:66 to 12:88). A comparison of the obtained yields and E/Z ratios of the products shown in Schemes 4 and [5](#page-5-0) revealed that, except for some cases, the efficacy of the bidentate directing group 4-

Scheme 5. Pd(II)-Catalyzed 4-Chloropicolinamide- and Chelation-Assisted Construction of Z-Cinnamylamines 4a−n^a

 a The E/Z ratios were determined from the NMR spectra of the corresponding crude reaction mixtures, and in most cases, the corresponding major isomer (Z-isomer) was isolated in pure form. ^bIn this case, compounds 4c' (E-isomer) and 4c (Z-isomer) were isolated in their pure forms. ^c15 mol $%$ of Pd(OAc)₂ was used. d_2 mmol of ArI was used.

chloro-2-picolinamide was comparable to that of 2-picolinamide.

To extend the substrate scope of this method, we attempted the arylation of N-(2-methylallyl)picolinamides 1c and 1d, which were derived from β -methylallylamine and the corresponding picolinic acids. Notably, N-(2-methylallyl) picolinamides 1c and 1d contain both γ -C(sp³)–H and γ - $C(sp^2)$ -H bonds, which can undergo the arylation under the experimental conditions. [Scheme 6](#page-6-0) shows the arylation of 1c and 1d with various aryl iodides under the optimized reaction conditions. We initially performed the Pd(II)-catalyzed reaction of 1c with aryl iodides containing a substituent at the para position (e.g., Me, Et, Cl, COOMe and i-Pr), which afforded the corresponding $γ$ -C(sp²)−H and $γ$ -C(sp³)−H bisarylated allylamine derivatives 5a−e with yields of 36−50% ([Scheme 6\)](#page-6-0).

We then performed the $Pd(II)$ -catalyzed reaction of $1c$ and 1d with various disubstituted aryl iodides, an aryl iodide containing a substituent at the meta position (e.g., COOEt), and heteroaryl iodides. These reactions also afforded the corresponding γ -C(sp²)–H and γ -C(sp³)–H bisarylated allylamine derivatives 5f−k and 7a with yields of 34−47%. It should be noted that the $Pd(II)$ -catalyzed reaction of 1c with 2-chloro-

4-iodopyridine afforded the γ -C(sp²)−H monoarylated product **6a** along with the expected $γ$ -C(sp²)-H and $γ$ -C(sp³)-H bisarylated allylamine derivative 5k. Given this interesting result, we performed the Pd(II)-catalyzed arylation of 1c and 1d with various iodopyridines, which afforded the corresponding γ -C(sp²)–H monoarylated products **6b–d** with yields of 40−41% [\(Scheme 6\)](#page-6-0).

Next, to elaborate our investigation on the $Pd(OAc)_{2}/$ AgOAc-catalyzed γ-C−H arylation of allylamines, we performed the Pd(II)-catalyzed reaction of picolinamide 1e with 2a, which gave derivative 8a with a yield of 40% with an E/Z ratio of 67:33 (eq 1, [Scheme 7](#page-6-0)). We then performed the Pd(II)-catalyzed reaction of N-allylpyrazine-2-carboxamide 1f with 2a, and this reaction afforded the γ-C−H arylated allylamine derivative 9a with a yield of only 35% and an E/Z ratio of 62:38 (eq 2, [Scheme 7](#page-6-0)). An initial attempt of the reaction of isoxazole-3-carboxamide $1g(1 \text{equiv})$ with p-tolyl iodide (1 equiv) in the presence of $Pd(OAc)₂$ (10 mol %) and AgOAc (2.2 equiv) in toluene at 110 °C afforded the γ-C−H arylated allylamine derivative 10a with a yield of 48% and an $E/$ Z ratio of 98:2 (entry 1, [Scheme 7](#page-6-0)). Surprised by this reaction, we then performed the arylation of 1g using different reaction Scheme 6. Pd(II)-Catalyzed Picolinamide- and Chelation-Assisted sp² and sp³ γ-C−H Arylation To Construct Z-Cinnamylamines 5a−k, 6a−d, and 7a

conditions to see whether we could obtain the Z-isomer as the major isomer. Accordingly, the reaction of $1g(1 \text{ equiv})$ with ptolyl iodide (1 equiv) in the presence of $Pd(OAc)$ ₂ (5 mol %) in toluene at rt afforded 10a in 33% yield with an E/Z ratio of 89:11 (entry 2, Scheme 7). The same reaction at 50 °C afforded 10a in an improved yield (78%) with an E/Z ratio of 98:2 (entry 3, Scheme 7). Apart from these reactions, other attempts to obtain the Z-isomer from the arylation of 1g under different reaction conditions were not fruitful (entries 4−7, Scheme 7). These results indicate that the arylation reactions comprising the substrates 1e−g underwent the conventional Heck-type reaction mechanism without chelation assistance by the corresponding directing groups (Scheme 7).^{31,[32,45a](#page-16-0)} We were interested in capitalizing on the reaction conditions that afforded compound 10a with E-stereochemistry (entry 3, Scheme 7) for synthesizing various γ-C−H arylated allylamine derivatives. Accordingly, treatment of 1g with various aryl iodides in the presence of $Pd(OAc)₂$ (5 mol %) in toluene at 50 °C afforded a series of γ-C−H arylated allylamines 10b−f with E-stereochemistry with yields of 60−75% (Scheme 7).

In the present work, the arylations of the picolinamide substrates 1a−d selectively afforded the corresponding γ-C−H

a The E/Z ratios were determined from the NMR spectra of the corresponding crude reaction mixtures. Compounds 10b−f were prepared using the conditions given in entry $3. \, b^b$ A complex mixture was obtained, and purification of the crude reaction mixture did not afford the corresponding cinnamylamine in pure form.

arylated products 3−7 with Z-stereochemistry as the predominant isomers [\(Table 1](#page-3-0), [Schemes 4](#page-4-0)−6). In concurrence with the generally proposed $Pd^{II}-Pd^{IV}$ catalytic cycle mechanism pertaining to the $Pd(OAc)_2/AgOAc\text{-}catalyzed,$ bidentate directing group assisted C−H functionaliza-tion,^{[9](#page-14-0)-[11](#page-15-0),[45](#page-16-0)} the observed Z-selective γ-C−H arylations of 1ad could be demonstrated via a plausible bidentate directing group assisted and chelation-controlled C−H activation mechanism [\(Scheme 8](#page-7-0)). In this process, it is believed that AgOAc helps in the ligand-exchange step to re-generate the Pd^H catalyst for the next catalytic cycle. $9-11$ $9-11$ $9-11$

Furthermore, the picolinamide substrate 1e and pyrazine-2 carboxamide substrate 1f were expected to afford the corresponding products 8a and 9a with Z-stereochemistry as the predominant isomers (Scheme 7); however, the corresponding products 8a and 9a with E-stereochemistry were obtained as the predominant isomers. These results are likely due to the absence of chelation assistance by the corresponding bidentate directing groups. Substrate 1e−g appears to only weakly coordinate with palladium, and therefore, the reactions Scheme 8. Plausible Mechanism for the Z-Selective γ-C−H Arylation of 1a−d

> bidentate directing group and chelation-assisted C-H functionalization

result in a mixture of products formed through two different mechanisms, such as a Heck-type mechanism $46-49$ leading to mechanisms, such as a Heck-type mechanism^{[46](#page-16-0)} the E-regioisomer and a bidentate directing group assisted, chelation-controlled mechanism leading to the Z-regioiso $mer.⁴⁵$

■ CONCLUSION

We investigated the Pd(II)-catalyzed, bidentate directing group- and chelation-assisted Z-selective γ-C(sp²)−H arylation of allylamines. The reactions of N-allylpicolinamides with various aryl and heteroaryl iodides in the presence of $Pd(OAc)$, and AgOAc led to the selective γ -C(sp²)−H arylation to construct various Z-cinnamylamine derivatives with low to high E/Z ratios (E/Z ratios up to 2:98). Additionally, the Pd(II)catalyzed arylation of an allylamine containing both γ -C(sp²)– H and γ -C(sp³)–H bonds afforded the γ -C(sp²)–H and γ -C(sp³)−H arylated cinnamylamine scaffolds. Although Hecktype γ-arylations of allylamines have generally afforded Ecinnamylamines, the present work demonstrated the construction of Z-cinnamylamine scaffolds with reasonably good E/Z ratios. The Pd(II)-catalyzed γ -C(sp²)–H arylation of Nallylpicolinamides was probed using different additives, directing groups and reaction conditions. In concurrence with the generally proposed mechanism pertaining to the Pd- (OAc)2/AgOAc-catalyzed, bidentate directing group-assisted C−H functionalization, the observed Z-selective γ -C(sp²)−H arylations of 1a−d were demonstrated via a plausible bidentate directing group and chelation-assisted C−H activation mechanism. Finally, the $Pd(OAc)₂/AgOAc-catalyzed$ arylation of N-allyl-5-methylisoxazole-3-carboxamide 1g was found to afford the E-cinnamylamine derivatives 10a−f, and it is assumed that the arylations of 1g occurred via a ligand-free Heck-type reaction mechanism to afford the corresponding Ecinnamylamine derivatives 10a−f.

EXPERIMENTAL SECTION

General Considerations. IR spectra of allylamines were recorded as thin films or KBr pellets. ${}^{1}H$ and ${}^{13}C$ NMR spectra of samples were recorded on 400 and 100 MHz spectrometers, respectively (using TMS as an internal standard). HRMS measurements reported in this work were obtained from QTOF mass analyzer using electrospray ionization method (ESI). Column chromatography was carried out using silica gel (100−200 mesh) or neutral alumina. Starting material preparation and C−H functionalization reactions were performed in anhydrous solvents under a nitrogen atmosphere wherever necessary. Isolated yields of all the compounds were reported, and yields were not optimized. TLC analysis was performed on silica gel or alumina plates, and the components were visualized by observation under iodine vapor. Amide starting materials used in the Pd(II)-catalyzed C− H arylation reactions were prepared (from their corresponding acids and amines) using the standard literature procedures.^{[9](#page-14-0)−[11](#page-15-0)} The E/Z ratios were determined from the NMR spectra of the corresponding crude reaction mixtures, and in the cases of the [Table 1](#page-3-0) and [Schemes](#page-4-0) [4,](#page-4-0) [5](#page-5-0) and [7](#page-6-0) the total yields of E/Z isomers were reported. With regard to the C−H arylation reactions of 1a−d, the data given here corresponds to the corresponding major isomers (Z-isomers) 3a−m, 4a−n, 5a−k, 7a and 6a−d. In some cases, the corresponding minor isomers (E-isomers) 3g′ and 4c′ were isolated and characterized. With regard to the C−H arylation reactions of 1e−g, the data given here correspond to the major isomers 8a, 9a, and 10a−f (E-isomers).

In concurrence with representative literature reports, $18c, d, 43, 45a$ the Z-stereochemistry of 3a−m and 4a−n (major isomers) and the Estereochemistry of 3g′ and 4c′ (minor isomers) were ascertained based on the observed characteristic coupling constant values of the corresponding doublet peaks of the olefin protons ($J = \sim 11.5$ Hz for the Z-isomers and $J = \sim 15.8$ Hz for the E-isomers). The Estereochemistry of 8a, 9a, and 10a−f were ascertained based on the observed characteristic coupling constant values of the corresponding doublet peaks of the olefin protons ($J = \sim 15.8$ Hz). Additionally, the observed E-stereochemistry of a representative compound 10e was confirmed from the X-ray structure analysis (see the [SI](http://pubs.acs.org/doi/suppl/10.1021/acs.joc.7b00535/suppl_file/jo7b00535_si_001.pdf) for the X-ray structure of 10e).

The following points are made with regard to separation of E/Z isomers. After arylation of the corresponding allylamines 1a,b, purification of the crude reaction mixture afforded the respective E/ Z cinnamylamines as a mixture since the corresponding E/Z cinnamylamines (3'/3 and 4'/4) had similar/close R_f values. Hence, the respective E/Z cinnamylamines were subjected to the repetitive column chromatographic purification to obtain the corresponding pure compounds. In some cases, we isolated the major isomers (Z-isomers) in pure form after the repetitive column chromatographic purification, and in some other cases, we got only a few column fractions of the corresponding major isomers (Z-isomers) in pure forms, which were used for characterization. Despite our repeated efforts to obtain the major isomers (Z-isomers) in pure forms, in some cases, the Z-isomers were obtained with traces of corresponding minor isomers due to similar R_f values of both major and minor isomers. With regard to isolation of minor isomers (E-isomers), except the minor isomers 3g′ and 4c′ (E-isomers), our repeated efforts to isolate the corresponding other E-isomers 3′ and 4′ in pure forms for characterization were not fruitful. The Pd(II)-catalyzed arylation of 1c,d afforded the corresponding double C−H arylated compounds 5a−k and 7a (Zisomers) as the major isomers. Though in some reactions the corresponding crude NMRs indicated the formation of trace amounts minor compounds, except the compound 6a, either we could not isolate any other corresponding minor isomers in pure forms or the reactions did not give any other isomers in characterizable amounts. The stereochemistry of 5a−k, 7a, and 6a−d was assigned based on the observed stereochemistry of 3a−m and 4a−n (major isomers).

Procedure for the Synthesis of N-Allylamides 1a−f,h−k,m. An oven-dried round-bottom flask (25 mL capacity) was charged with an appropriate carboxylic acid (1.2 mmol, 1 equiv), anhydrous DCM (4−6 mL), and two to three drops of DMF. To this solution was added oxalyl chloride (1.5 mmol, 1.5 equiv, 190 mg) dropwise at 0 $^{\circ}$ C.

The mixture was stirred at rt for 12 h, and then the solvent was removed in vacuo and dissolved in DCM (4−6 mL). The resulting acid chloride solution was immediately used in the next step without further purification. Another oven-dried round-bottom flask (25 mL capacity) was charged with an appropriate allylamine (1.0 mmol, 1 equiv), $Et₃N$ (1.5 mmol, 1.5 equiv, 152 mg), and DMAP (0.1 mmol, 0.1 equiv, 12 mg). To this solution was added the acid chloride solution (obtained in the previous step) dropwise at 0 °C, and after the addition the solution was warmed to rt and allow to stir overnight. Then the reaction mixture was quenched with satd aq $NAHCO₃$ solution (10−15 mL), and the organic layer was separated, dried over anhydrous $MgSO_4$, and evaporated in vacuo. The crude reaction mixture was purified by column chromatography on silica gel (eluent: EtOAc/hexanes = 30:70) to afford the corresponding carboxamides 1a−f,h−k,m.

Procedure for the Synthesis of Carboxamides 1g and 1l. An oven-dried round-bottom flask (25 mL capacity) was charged with 5 methylisoxazole-3-carboxylic acid (1 mmol, 1 equiv) and DCM (6 mL) under a nitrogen atm. EDCI (1.1 mmol, 1.1 equiv, 172 mg) and HOBt·H₂O (1.1 mmol, 1.1 equiv, 168 mg) were added dropwise at 0 °C, and the reaction mixture was stirred for 15 min. Then to the reaction mixture was added an appropriate allylamine (1 mmol, 1.1 equiv) dropwise at 0 °C. The solution was warmed to room temperature, and stirring was continued for 12 h. After this period, water (4−7 mL) was added and the solution extracted with DCM (4− 7 mL, 2−3 times). The combined organic layers were washed with satd ag NaHCO₃ (10 mL) and dried over anhydrous $MgSO₄$, and the solvent was evaporated in vacuo. The crude reaction mixture was purified by column chromatography on silica gel (eluent: EtOAc/

hexanes = 35:65) to afford the corresponding products 1g and 1l.
General Procedure for the Pd(II)-Catalyzed Arylation of N-Allylamide Derivatives 1a-m and the Preparation of 3a-m/ 4a−n/5a−k/6a−d/7a/10a−f. An oven-dried round-bottom flask (10 mL capacity) was charged with an appropriate N-allylamide derivative (0.25 mmol, 1 equiv), an appropriate aryl iodide (1.0 mmol, 4.0 equiv), $Pd(OAc)_2$ (10 mol %, 5.6 mg,), AgOAc (0.55 mmol, 2.2 equiv, 91.8 mg), and toluene (3 mL). This reaction mixture was heated at 110 °C for 24−48 h under a nitrogen atm. After this period, the reaction mixture was concentrated in vacuo, and purification of the resulting reaction mixture by column chromatography on silica gel furnished the corresponding γ-C−H arylated allylamine derivatives 3a−m/4a−n/5a−k/6a−d/7a/10a−f (see the corresponding tables and schemes for specific examples).

N-Allylpicolinamide (1a).^{[50a](#page-17-0)} Compound 1a was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a brown liquid: R_f = 0.51 (EtOAc/hexane = 1:4); yield 77% (126 mg); IR (DCM) 3446, 2064, 1642, 1529, 1465 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.55 (dd, 1H, J₁ = 4.8 Hz, J₂ = 0.8 Hz), 8.21 (dd, 1H, $J_1 = 7.8$ Hz, $J_2 = 1.0$ Hz), 8.17 (br s, 1H), 7.85 (t, 1H, $J =$ 7.7 Hz), 7.45−7.41 (m, 1H), 5.99−5.89 (m, 1H), 5.30−5.16 (m, 2H), 4.13−4.10 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.7, 148.0, 137.3, 134.0, 126.2, 122.2, 116.3, 41.7; HRMS (ESI) m/z [M + $[H]^+$ calcd for $C_9H_{11}N_2O$ 163.0871, found 163.0875.

N-Allyl-4-chloropicolinamide (1b). 50b 50b 50b Compound 1b was obtained after purification by column chromatography on silica gel (EtOAc/ hexanes = 30:70) as a pale yellow liquid: R_f = 0.50 (EtOAc/hexane = 1:4); yield 67% (133 mg); IR (DCM) 3441, 2063, 1643, 1524, 1290 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.41 (d, 1H, J = 5.2 Hz), 8.17 (d, 1H, J = 2.0 Hz), 8.10 (br s, 1H), 7.41 (td, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 5.95−5.85 (m, 1H), 5.26−5.13 (m, 2H), 4.09−4.06 (m, 2H); 13C NMR (100 MHz, CDCl₃) δ 163.0, 151.3, 149.0, 145.8, 133.8, 126.3, 122.9, 116.6, 41.9; HRMS (ESI) m/z [M + H]⁺ calcd for C₉H₁₀ClN₂O 197.0482, found 197.0484.

N-(2-Methylallyl)picolinamide (1c). Compound 1c was obtained after purification by column chromatography on silica gel (EtOAc/ hexanes = 30:70) as a colorless liquid: R_f = 0.52 (EtOAc/hexane = 1:4); yield 69% (123 mg); IR (DCM) 3389, 1675, 1527, 1289 cm⁻¹;
¹H NMR (400 MHz, CDCL) δ 8 32–8 31 (m, 1H) 8 17 (br.s. 1H) ¹H NMR (400 MHz, CDCl₃) δ 8.32–8.31 (m, 1H), 8.17 (br s, 1H), 8.00 (dd, 1H, $J_1 = 7.8$ Hz, $J_2 = 0.9$ Hz), 7.61 (td, 1H, $J = 7.7$ Hz, $J = 1.8$ Hz), 7.21−7.19 (m, 1H), 4.69−4.63 (m, 2H), 3.82 (d, 2H, J = 6.4 Hz),

1.55 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.1, 149.6, 147.9, 141.6, 137.2, 126.1, 122.1, 110.8, 44.7, 20.2; HRMS (ESI) m/z [M + $[H]^+$ calcd for $C_{10}H_{13}N_2O$ 177.1028, found 177.1023.

4-Chloro-N-(2-methylallyl)picolinamide (1d). Compound 1d was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/ hexanes = 1:4); yield 35% (74 mg); IR (DCM) 3391, 1677, 1527, 1292 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.44 (d, 1H, J = 5.2 Hz), 8.21 (d, 1H, J = 1.4 Hz), 8.14 (br s, 1H), 7.44–7.42 (m, 1H), 4.90 (s, 1H), 4.87 (s, 1H), 4.01 (d, 2H, J = 6.2 Hz), 1.78 (s, 3H); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3)$ δ 163.1, 151.3, 149.0, 145.9, 141.6, 126.3, 123.0, 111.2, 45.1, 20.4; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₀H₁₂ClN₂O 211.0638, found 211.0629.

 N_2 , N_6 -Diallylpyridine-2,6-dicarboxamide (1e). Compound 1e was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.52$ (EtOAc/ hexanes = 1:4); yield 65% (161 mg); IR (DCM) 3287, 1667, 1537, 1445 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 9.18 (t, 2H, J = 6.0 Hz), 8.28 (d, 2H, J = 7.8 Hz), 7.95 (t, 1H, J = 7.8 Hz), 5.69–5.59 (m, 2H), 4.96−4.91 (m, 2H), 4.83−4.81 (m, 2H), 3.86 (t, 4H, $J = 5.8$ Hz); ¹³C NMR (100 MHz, CDCl₃) δ 164.0, 148.8, 138.9, 133.8, 124.9, 116.1, 42.0; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₃H₁₅N₃NaO₂ 268.1062, found 268.1068.

N-Allylpyrazine-2-carboxamide (1f). Compound 1f was obtained after purification by column chromatography on silica gel (EtOAc/ hexanes = 40:60) as a colorless liquid: R_f = 0.50 (EtOAc/hexanes = 1:4); yield 70% (115 mg); IR (DCM) 3397, 1667, 1532, 1402 cm⁻¹;
¹H NMR (400 MHz, CDCl) 8.9.41 (dd. 1H I – 3.4 Hz, I – 1.3 Hz) ¹H NMR (400 MHz, CDCl₃) δ 9.41 (dd, 1H, J₁ = 3.4 Hz, J₂ = 1.3 Hz), 8.74 (dd, 1H, $J_1 = 5.1$ Hz, $J_2 = 2.5$ Hz), 8.53 (br s, 1H), 7.95 (br s, 1H), 5.97−5.88 (m, 1H), 5.30−5.16 (m, 2H), 4.13−4.10 (m, 2H); 13C NMR (100 MHz, CDCl₃) δ 162.8, 147.3, 144.4, 144.4, 142.6, 133.6, 116.8, 41.8; HRMS (ESI) m/z [M + H]⁺ calcd for C₈H₁₀N₃O 164.0824, found 164.0822.

N-Allyl-5-methylisoxazole-3-carboxamide (1g). Compound 1g was obtained after purification by column chromatography on neutral alumina (EtOAc/hexanes = 35:65) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexane = 1:4); yield 50% (83 mg); IR (DCM) 3332, 1673, 1599, 1458, 1280 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.07 (br s, 1H), 6.43 (s, 1H), 5.92−5.83 (m, 1H), 5.26−5.14 (m, 2H), 4.04 (t, 2H, J = 5.8 Hz), 2.45 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 159.0, 158.7, 133.4, 116.8, 101.4, 41.7, 12.3; HRMS (ESI) m/z [M + H ⁺ calcd for C₈H₁₁N₂O₂ 167.0821, found 167.0815.

N-Allyl-1-naphthamide (1h). Compound 1h was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.52 (EtOAc/hexanes = 1:4); yield 65% (138 mg); IR (KBr) 3284, 1640, 1536, 1422 cm[−]¹ ; 1 H NMR (400 MHz, CDCl₃) δ 8.13 (d, 1H, J = 8.1 Hz), 7.77–7.73 (m, 2H), 7.45– 7.37 (m, 2H), 7.32 (d, 1H, $J = 7.0$ Hz), 7.18 (t, 1H, $J = 7.9$ Hz), 6.99 (br s, 1H), 5.80−5.71 (m, 1H), 5.12−5.01 (m, 2H), 3.84 (t, 2H, J = 5.6 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 169.6, 139.1, 134.1, 133.5, 130.3, 130.1, 128.2, 126.8, 126.2, 125.5, 125.0, 124.6, 116.1, 42.2; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₄H₁₄NO 212.1075, found 212.1082.

N-Allyl-3-phenylpropanamide (1i).^{[50c](#page-17-0)} Compound 1i was obtained after purification by column chromatography on silica gel (EtOAc/ hexanes = 30:70) as a colorless liquid: R_f = 0.51 (EtOAc/hexanes = 1:4); yield 72% (137 mg); IR (DCM) 3293, 1643, 1551, 1454, 1262 cm[−]¹ ; 1 H NMR (400 MHz, CDCl3) δ 7.30−7.26 (m, 2H), 7.22−7.19 (m, 3H), 6.28 (br s, 1H), 5.82−5.72 (m, 1H), 5.10−5.06 (m, 2H), 3.83 (t, 2H, J = 5.6 Hz), 2.97 (t, 2H, J = 7.5 Hz), 2.52 (t, 2H, J = 7.5 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 172.3, 140.9, 134.2, 128.5, 128.3, 126.2, 116.1, 41.9, 38.3, 31.8; HRMS (ESI) m/z [M + H]⁺ calcd for C12H16NO 190.1232, found 190.1228.

N-(2-Methylallyl)-3-phenylpropanamide (1**j**).^{[50d](#page-17-0)} Compound 1j was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.50 (EtOAc/ hexane = 1:4); yield 74% (152 mg); IR (DCM) 3294, 1648, 1553, 1454 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.30−7.27 (m, 2H), 7.22− 7.20 (m, 3H), 6.00 (br s, 1H), 4.78 (s, 1H), 4.70 (s, 1H), 3.77 (d, 2H, $J = 5.9$ Hz), 2.99 (t, 2H, $J = 7.9$ Hz), 2.54 (t, 2H, $J = 7.9$ Hz), 1.67 (s,

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3H); ¹³C NMR (100 MHz, CDCl₃) δ 172.2, 141.9, 140.9, 128.5, 128.4, 126.2, 110.8, 45.0, 38.4, 31.8, 20.3; HRMS (ESI) m/z [M + H]⁺ calcd for $C_{13}H_{18}NO$ 204.1388, found 204.1379.

N-(2-(Cyclohex-1-en-1-yl)ethyl)picolinamide (1k). ^{[50e](#page-17-0)} Compound 1k was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 40:60) as a colorless liquid: $R_f = 0.50$ (EtOAc/ hexanes = 1:4); yield 80% (184 mg); IR (DCM) 3392, 1665, 1590, 1433 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.48 (dd, 1H, J₁ = 4.7 Hz, $J_1 = 0.5$ Hz), 8.14 (d, 1H, J = 7.8 Hz), 8.05 (br s, 1H), 7.77 (td, 1H, J = 7.7 Hz, J = 1.7 Hz), 7.37−7.33 (m, 1H), 5.47 (s, 1H), 3.52−3.47 (m, 2H), 2.22−2.19 (m, 2H), 1.94−1.92 (m, 4H), 1.60−1.48 (m, 4H); 13C NMR (100 MHz, CDCl₃) δ 164.1, 150.0, 148.0, 137.2, 134.5, 125.9, 123.4, 122.0, 37.7, 37.5, 28.0, 25.2, 22.8, 22.3; HRMS (ESI) m/z [M + H ⁺ calcd for C₁₄H₁₉N₂O 231.1497, found 231.1491.

N-(2-(Cyclohex-1-en-1-yl)ethyl)-5-methylisoxazole-3-carboxamide (1I). Compound 11 was obtained after purification by column chromatography on neutral alumina (EtOAc/hexanes = 35:65) as a colorless liquid: $R_f = 0.52$ (EtOAc/hexanes = 1:4); yield 65% (153) mg); IR (DCM) 3365, 1660, 1601, 1545, 1270 cm^{−1};¹H NMR (400 MHz, CDCl₃) δ 6.83 (br s, 1H), 6.42 (s, 1H), 5.51 (s, 1H), 3.52–3.47 $(m, 2H)$, 2.47 (s, 3H), 2.22 (t, 2H, $I = 6.8$ Hz), 1.99–1.95 (m, 4H), 1.65−1.53 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 171.0, 159.0, 158.9, 134.2, 123.9, 101.4, 37.4, 37.3, 27.9, 25.2, 22.8, 22.3, 12.3; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₃H₁₉N₂O₂ 235.1447, found 235.1440.

(E)-N-(Quinolin-8-yl)hex-3-enamide (1m). Compound 1m was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a brown liquid: $R_f = 0.51$ (EtOAc/ hexane = 1:4); yield 70% (168 mg); IR (DCM) 3054, 2305, 1422, 1265 cm[−]¹ ; 1 H NMR (400 MHz, CDCl3) δ 10.10 (br s, 1H), 8.80− 8.77 (m, 2H), 8.13 (dd, 1H, $J_1 = 8.3$ Hz, $J_2 = 1.6$ Hz), 7.53 (dd, 1H, J_1 $= 8.2$ Hz, $J_2 = 7.7$ Hz), 7.48 (dd, 1H, $J_1 = 8.2$ Hz, $J_2 = 1.6$ Hz), 7.43 (dd, 1H, $J_1 = 8.3$ Hz, $J_2 = 4.2$ Hz), $5.92 - 5.72$ (m, 2H), 3.28 (d, 2H, $J =$ 7.1 Hz), 2.24−2.16 (m, 2H), 1.14 (t, 3H, J = 7.4 Hz); 13C NMR (100 MHz, CDCl₃) δ 170.2, 148.1, 138.7, 138.5, 136.3, 134.5, 127.9, 127.4, 121.6, 121.5, 121.4, 116.3, 42.1, 25.8, 13.6; HRMS (ESI) m/z [M + $[H]^+$ calcd for $C_{15}H_{17}N_2O$ 241.1341, found 241.1330.

(Z)-N-(3-(4-Methoxyphenyl)allyl)picolinamide (3a). Compound 3a (2) major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 25:75) as a brown liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 64% (43 mg, $E/Z =$ 11:89); IR (DCM) 3441, 1667, 1511, 1251, 1032 cm⁻¹; ¹H NMR (400 MHz, CDCl3) δ 8.57−8.55 (m, 1H), 8.24−8.22 (m, 1H), 8.16 (br s, 1H), 7.87 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.46–7.43 (m, 1H), 7.26 (d, 2H, J = 8.6 Hz), 6.92 (d, 2H, J = 8.8 Hz), 6.59 (d, 1H, J = 11.5 Hz), 5.71 (dt, 1H, $J_1 = 11.5$ Hz, $J_2 = 6.7$ Hz), 4.41 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.8$ Hz), 3.84 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 158.8, 149.8, 148.1, 137.4, 131.3, 130.1, 129.0, 126.2, 122.3, 113.8, 55.3, 37.9; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₆H₁₆N₂NaO₂ 291.1109, found 291.1099.

 (Z) -N-(3-Phenylallyl)picolinamide (3b). Compound 3b $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a yellow liquid: $R_f = 0.51$ (EtOAc/hexane = 1:4); yield 57% (34 mg, $E/Z = 25:75$); IR (DCM) 3441, 1659, 1524, 1383, 1041 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.57−8.55 (m, 1H), 8.25−8.22 (m, 1H), 8.17 (br s, 1H), 7.87 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.46–7.43 (m, 1H), 7.41–7.37 (m, 2H), 7.33−7.28 (m, 3H), 6.67 (d, 1H, J = 11.6 Hz), 5.81 (dt, 1H, J₁ = 11.6 Hz, $J_2 = 6.7$ Hz), 4.41 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.87$ Hz); ¹³C NMR (100 MHz, CDCl3) δ 164.2, 149.8, 148.1, 137.4, 136.4, 131.8, 128.8, 128.4, 127.9, 127.3, 126.2, 122.3, 37.8; HRMS (ESI) m/z [M + Na]+ calcd for $C_{15}H_{14}N_2NaO$ 261.1004, found 261.0995.

(Z)-N-(3-(4-Ethylphenyl)allyl)picolinamide (3c). Compound 3c $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 66% (44 mg, $E/Z =$ 25:75); IR (DCM) 3441, 1735, 1623, 1383, 1046 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.57–8.55 (m, 1H), 8.25–8.22 (m, 1H), 8.16 (br s, 1H), 7.88 (td, 1H, J₁ = 7.7 Hz, J₂ = 1.7 Hz), 7.47–7.43 (m, 1H), 7.26−7.20 (m, 4H), 6.63 (d, 1H, J = 11.6 Hz), 5.76 (dt, 1H, J₁ = 11.6

Hz, $J_2 = 6.7$ Hz), 4.42 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.8$ Hz), 2.67 (q, 2H, J $= 7.6$ Hz), 1.26 (t, 3H, J = 7.6 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.8, 148.1, 143.5, 137.4, 133.7, 131.8, 128.8, 127.9, 127.1, 126.2, 122.3, 37.9, 28.6, 15.6; HRMS (ESI) m/z [M + Na]⁺ calcd for $C_{17}H_{18}N_2NaO$ 289.1317, found 289.1309.

(Z)-N-(3-(4-Pentylphenyl)allyl)picolinamide (3d). Compound 3d $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 69% (53 mg, $E/Z =$ 9:91); IŘ (DCM) 3442, 2921, 1643, 1390 cm^{−1}; ¹H NMR (400 MHz, CDCl₃) δ 8.56 (d, 1H, J = 4.7 Hz), 8.23 (d, 1H, J = 7.8 Hz), 8.15 (br s, 1H), 7.87 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.46–7.43 (m, 1H), 7.23 $(d, 2H, J = 8.2 \text{ Hz})$, 7.20 $(d, 2H, J = 8.2 \text{ Hz})$, 6.63 $(d, 1H, J = 11.6 \text{ Hz})$ Hz), 5.76 (dt, 1H, $J_1 = 11.6$ Hz, $J_2 = 6.7$ Hz), 4.42 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.7$ Hz), 2.62 (t, 2H, J = 7.6 Hz), 1.67–1.60 (m, 2H), 1.36–1.27 (m, 4H), 0.92 (t, 3H, J = 6.8 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.9, 148.1, 142.2, 137.4, 133.7, 131.8, 128.8, 128.4, 127.1, 126.2, 122.3, 37.9, 35.7, 31.5, 31.2, 22.6, 14.1; HRMS (ESI) m/z [M + Na]⁺ calcd for $C_{20}H_{24}N_2N_3$ NaO 331.1786, found 331.1776.

 $(Z)-N-(3-(4-Hexylphenyl)$ allyl)picolinamide (3e). Compound 3e ((Z) major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 64% (52 mg, $E/Z =$ 14:86); IR (DCM) 2928, 1678, 1523, 1434 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.57–8.55 (m, 1H), 8.23 (d, 1H, J = 7.8 Hz), 8.15 (br s, 1H), 7.87 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.46–7.43 (m, 1H), 7.23 (d, 2H, J = 8.2 Hz), 7.20 (d, 2H, J = 8.2 Hz), 6.64 (d, 1H, J = 11.6 Hz), 5.76 (dt, 1H, $J_1 = 11.6$ Hz, $J_2 = 6.7$ Hz), 4.42 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.7 \text{ Hz}$), 2.62 (t, 2H, J = 7.6 Hz), 1.68–1.59 (m, 2H), 1.38–1.27 (m, 6H), 0.91 (t, 3H, $J = 6.6$ Hz); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.9, 148.1, 142.2, 137.4, 133.7, 131.8, 128.8, 128.4, 127.1, 126.2, 122.3, 37.9, 35.7, 31.8, 31.4, 29.0, 22.6, 14.1; HRMS (ESI) m/z $[M + H]^{+}$ calcd for $C_{21}H_{27}N_{2}O$ 323.2123, found 323.2111.

(Z)-N-(3-(4-Isopropylphenyl)allyl)picolinamide (3f). Compound 3f ((Z) major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 50% (35 mg, $E/Z =$ 20:80); IR (DCM) 3390, 1672, 1523, 1464 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.57–8.55 (m, 1H), 8.23 (d, 1H, J = 7.8 Hz), 8.15 (br s, 1H), 7.87 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.46–7.43 (m, 1H), 7.25 (s, 4H), 6.63 (d, 1H, J = 11.6 Hz), 5.76 (dt, 1H, J₁ = 11.6 Hz, J₂ = 6.8 Hz), 4.42 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.8$ Hz), 2.97–2.90 (m, 1H), 1.27 (d, 6H, J = 6.9 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.9, 148.1, 137.4, 133.9, 131.8, 128.8, 127.1, 126.4, 126.2, 122.3, 37.9, 33.9, 24.0; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₈H₂₀N₂NaO 303.1473, found 303.1462.

(E)-N-(3-(p-Tolyl)allyl)picolinamide $(3g')$. Compound $3g'$ $((E)$ minor isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a yellow liquid: R_f = 0.51 (EtOAc/hexane = 1:4); yield 11% (7 mg, E/Z = 29:71); IR (DCM) 3442, 1738, 1642, 1365, 1216 cm[−]¹ ; 1 H NMR (400 MHz, CDCl₃) δ 8.59–8.57 (m, 1H), 8.26–8.24 (m, 1H), 8.21 (br s, 1H), 7.88 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.47–7.44 (m, 1H), 7.30 (d, 2H, $J = 8.8$ Hz), 7.13 (d, 2H, $J = 8.8$ Hz), 6.61 (d, 1H, $J = 15.8$ Hz), 6.27 (dt, 1H, $J_1 = 15.8$ Hz, $J_2 = 6.3$ Hz), 4.28 (td, 2H, $J_1 = 6.3$ Hz, $J_2 =$ 1.4 Hz), 2.35 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.9, 148.1, 137.5, 137.4, 133.8, 132.2, 129.3, 126.3, 126.2, 124.3, 122.3, 41.5, 21.2; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₆H₁₆N₂NaO 275.1160, found 275.1147.

 (Z) -N-(3-(p-Tolyl)allyl)picolinamide (3g). Compound 3g $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.51 (EtOAc/hexanes = 1:4); yield 71% (45 mg, E/Z = 29:71); IR (DCM) 3386, 1668, 1525, 1463, 1043 cm[−]¹ ; 1 H NMR (400 MHz, CDCl₃) δ 8.57–8.55 (m, 1H), 8.25–8.22 (m, 1H), 8.16 (br s, 1H), 7.87 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.46–7.43 (m, 1H), 7.23–7.12 $(m, 4H)$, 6.63 (d, 1H, J = 11.5 Hz), 5.75 (dt, 1H, J₁ = 11.5 Hz, J₂ = 6.7 Hz), 4.41 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.8$ Hz), 2.38 (s, 3H); ¹³C NMR (100 MHz, CDCl3) δ 164.2, 149.8, 148.1, 137.4, 137.1, 133.5, 131.7,

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129.1, 128.8, 127.2, 126.2, 122.3, 37.9, 21.2; HRMS (ESI) m/z [M + Na]⁺ calcd for $C_{16}H_{16}N_2NaO$ 275.1160, found 275.1148.

(Z)-N-(3-(3,4-Dimethylphenyl)allyl)picolinamide (3h). Compound 3h (2) major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 40% (27 mg, $E/Z =$ 29:71); IR (DCM) 3441, 1643, 1367, 1232 cm^{−1}; ¹H NMR (400 MHz, CDCl₃) δ 8.57–8.55 (m, 1H), 8.25–8.22 (m, 1H), 8.15 (br s, 1H), 7.87 (td, 1H, J¹ = 7.6 Hz, J² = 1.7 Hz), 7.46−7.43 (m, 1H), 7.15 (d, 1H, $J = 7.6$ Hz), $7.08 - 7.05$ (m, 2H), 6.60 (d, 1H, $J = 11.6$ Hz), 5.74 (dt, 1H, $J_1 = 11.6$ Hz, $J_2 = 6.7$ Hz), 4.42 (td, 2H, $J_1 = 6.7$ Hz, $J_2 =$ 1.8 Hz), 2.30 (s, 3H), 2.29 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.9, 148.1, 137.4, 136.5, 135.8, 134.0, 131.8, 130.1, 129.6, 127.0, 126.3, 126.2, 122.3, 37.9, 19.9, 19.5; HRMS (ESI) m/z [M + Na]⁺ calcd for $C_{17}H_{18}N_2N_4O$ 289.1317, found 289.1308.

(Z)-N-(3-(2,3-Dihydrobenzo[b][1,4]dioxin-6-yl)allyl)picolinamide (3i). Compound 3i $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a pale yellow liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 59% (44 mg, $E/Z = 27:73$); IR (DCM) 3442, 1671, 1506, 1433 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.56–8.55 (m, 1H), 8.23 (d, 1H, J = 7.8 Hz), 8.16 (br s, 1H), 7.86 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.45–7.42 (m, 1H), 6.87 (d, 1H, J = 8.3 Hz), 6.83 (d, 1H, J = 2.0 Hz), 6.80 (dd, 1H, $J_1 = 8.3$ Hz, $J_2 = 2.0$ Hz), 6.53 (d, 1H, $J = 11.5$ Hz), 5.70 (dt, 1H, $J_1 =$ 11.6 Hz, J_2 = 6.7 Hz), 4.40 (td, 2H, J_1 = 6.6 Hz, J_2 = 1.8 Hz), 4.29 (s, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.8, 148.1, 143.2, 142.9, 137.4, 131.1, 130.0, 126.8, 126.2, 122.3, 122.2, 117.6, 117.1, 64.4, 64.3, 37.9; HRMS (ESI) m/z $[M + H]^+$ calcd for $C_{17}H_{17}N_2O_3$: 297.1239, found 297.1248.

(Z)-N-(3-(3,5-Dimethylphenyl)allyl)picolinamide (3j). Compound $3j$ $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 51% (34 mg, $E/Z =$ 38:62); IR (DCM) 1717, 1679, 1521, 1203 cm⁻¹; ¹H NMR (400 MHz, CDCl3) δ 8.57−8.55 (m, 1H), 8.25−8.22 (m, 1H), 8.15 (br s, 1H), 7.87 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.46–7.43 (m, 1H), 6.94– 6.93 (m, 3H), 6.60 (d, 1H, J = 11.6 Hz), 5.76 (dt, 1H, J₁ = 11.6 Hz, J₂ $= 6.7$ Hz), 4.42 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.9$ Hz), 2.35 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.9, 148.1, 137.9, 137.4, 136.3, 132.0, 129.0, 127.6, 126.6, 126.2, 122.3, 37.9, 21.4; HRMS (ESI) m/z $[M + Na]^{+}$ calcd for $C_{17}H_{18}N_2N_3N_4O$ 289.1317, found 289.1317.

(Z)-Ethyl 3-(3-(Picolinamido)prop-1-en-1-yl)benzoate (3k). Compound 3k ((Z) major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a yellow liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 54% (42 mg, E/ $Z = 21:79$); IR (DCM) 3380, 1717, 1523, 1282 cm⁻¹;¹H NMR (400 MHz, CDCl₃) δ 8.56–8.55 (m, 1H), 8.22 (d, 1H, J = 7.8 Hz), 8.19 (br s, 1H), 7.97 (dd, 2H, $J_1 = 6.4$ Hz, $J_2 = 1.4$ Hz), 7.87 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.53–7.42 (m, 3H), 6.68 (d, 1H, J = 11.6 Hz), 5.87 (dt, 1H, $J_1 = 11.6$ Hz, $J_2 = 6.7$ Hz), 4.43–4.36 (m, 4H), 1.41 (t, 3H, J = 7.1 Hz); 13 C NMR (100 MHz, CDCl₃) δ 166.4, 164.2, 149.7, 148.1, 137.4, 136.6, 133.0, 130.9, 130.6, 129.8, 129.2, 128.5, 128.4, 126.3, 122.3, 61.1, 37.7, 14.3; HRMS (ESI) m/z $[M + H]^+$ calcd for $C_{18}H_{19}N_2O_3$ 311.1396, found 311.1410.

(Z)-N-(3-(Thiophene-2-yl)allyl)picolinamide (3l). Compound 3l ((Z) major isomer)was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a pale yellow liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 69% (42 mg, E/ $Z = 10.90$); IR (DCM) 3442, 1662, 1532, 1386 cm⁻¹;¹H NMR (400 MHz, CDCl3) δ 8.57−8.55 (m, 1H), 8.24 (d, 1H, J = 7.8 Hz), 8.24 (br s, 1H), 7.87 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.46–7.43 (m, 1H), 7.34 (t, 1H, $J = 3.0$ Hz), 7.06 (t, 2H, $J = 3.5$ Hz), 6.70 (d, 1H, $J = 11.6$ Hz), 5.71 (dt, 1H, $J_1 = 11.6$ Hz, $J_2 = 6.7$ Hz), 4.51 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.9$ Hz); ¹³C NMR (100 MHz, CDCl₃) δ 164.3, 149.8, 148.1, 139.3, 137.4, 128.1, 127.2, 126.2, 126.1, 126.1, 124.0, 122.3, 38.3; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₃H₁₃N₂OS 245.0749, found 245.0749.

(Z)-N-(3-(5-Bromopyridin-2-yl)allyl)picolinamide (3m). Compound $3m$ $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a

pale yellow liquid: $R_f = 0.45$ (EtOAc/hexanes = 1:4); yield 50% (40 mg, E/Z = 2:98); IR (DCM) 3337, 1651, 1521 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.73 (d, 1H, J = 2.2 Hz), 8.67 (br s, 1H), 8.58–8.56 (m, 1H), 8.22 (d, 1H, J = 7.8 Hz), 7.86 (td, 1H, J₁ = 7.7 Hz, J₂ = 1.7 Hz), 7.80 (dd, 1H, $J_1 = 8.4$ Hz, $J_2 = 2.4$ Hz), 7.45–7.41 (m, 1H), 7.15 (d, 1H, $J = 8.3$ Hz), 6.50 (d, 1H, $J = 11.7$ Hz), 6.09 (dt, 1H, $J_1 = 11.6$ Hz, $J_2 = 6.7$ Hz), 4.66 (td, 2H, $J_1 = 6.5$ Hz, $J_2 = 1.7$ Hz); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3)$ δ 164.3, 154.1, 150.4, 150.1, 148.1, 138.9, 137.3, 133.7, 129.0, 126.1, 125.5, 122.3, 118.7, 37.9; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₄H₁₃BrN₃O 318.0242, found 318.0244.

(Z)-4-Chloro-N-(3-phenylallyl)picolinamide (4a). Compound 4a ((Z) major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 50% (34 mg, $E/Z =$ 34:66); IR (DCM) 3382, 1674, 1525, 1262 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.45 (d, 1H, J = 5.2 Hz), 8.24 (d, 1H, J = 1.9 Hz), 8.08 (br s, 1H), 7.46 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 7.41–7.37 (m, 2H), 7.32–7.28 (m, 3H), 6.68 (d, 1H, J = 11.6 Hz), 5.79 (dt, 1H, J₁ = 11.5 Hz, $J_2 = 6.8$ Hz), 4.41 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.8$ Hz); ¹³C NMR (100 MHz, CDCl₃) δ 163.0, 151.3, 149.0, 145.9, 136.3, 132.1, 128.8, 128.4, 127.5, 127.4, 126.4, 122.9, 37.9; HRMS (ESI) m/z [M + $[H]^+$ calcd for $C_{15}H_{14}CIN_2O$ 273.0795, found 273.0782.

(Z)-4-Chloro-N-(3-(4-methoxyphenyl)allyl)picolinamide (4b). Compound $4b$ $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a yellow liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 41% (31 mg, $E/Z = 29:71$); IR (DCM) 3442, 1670, 1511, 1383 cm⁻¹; ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3)$ δ 8.45 (d, 1H, J = 5.2 Hz), 8.24 (d, 1H, J = 2.0 Hz), 8.07 (br s, 1H), 7.45 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 7.25 (d, 2H, $J = 8.6$ Hz), 6.92 (d, 2H, $J = 8.8$ Hz), 6.60 (d, 1H, $J = 11.5$ Hz), 5.69 (dt, 1H, $J_1 = 11.5$ Hz, $J_2 = 6.7$ Hz), 4.40 (td, 2H, $J_1 = 6.7$ Hz, $J_2 =$ 1.8 Hz), 3.84 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 163.0, 158.8, 151.3, 149.0, 145.9, 131.6, 130.1, 128.9, 126.4, 125.8, 122.9, 113.8, 55.3, 38.0; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₆H₁₆ClN₂O₂ 303.0900, found 303.0889.

(E)-4-Chloro-N-(3-(4-isopropylphenyl)allyl)picolinamide (4c′). Compound $4c'$ ((E) minor isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a yellow liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 8% (7 mg, E/ $Z = 19:81$); IR (DCM) 3412, 1670, 1520, 1386 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.47 (d, 1H, J = 5.2 Hz), 8.25 (d, 1H, J = 2.0 Hz), 8.12 (d, 1H, J = 0.5 Hz), 7.46 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 7.33 $(d, 2H, J = 8.2 \text{ Hz})$, 7.19 $(d, 2H, J = 8.2 \text{ Hz})$, 6.61 $(d, 1H, J = 15.8 \text{ Hz})$ Hz), 6.25 (dt, 1H, $J_1 = 15.8$ Hz, $J_2 = 6.4$ Hz), 4.27 (t, 2H, $J = 6.0$ Hz), 2.94−2.87 (m, 1H), 1.25 (d, 6H, J = 6.9 Hz); ¹³C NMR (100 MHz, CDCl3) δ 163.0, 151.4, 149.0, 148.7, 145.9, 134.1, 132.4, 126.7, 126.4, 126.4, 124.1, 123.0, 41.6, 33.9, 23.9; HRMS (ESI) m/z [M + H]⁺ calcd for $C_{18}H_{20}C/N_2O$ 315.1264, found 315.1266.

(Z)-4-Chloro-N-(3-(4-isopropylphenyl)allyl)picolinamide (4c). Compound $4c$ $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a yellow liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 53% (42 mg, $E/Z = 19.81$); IR (DCM) 3385, 1673, 1532, 1266 cm⁻¹; ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3)$ δ 8.45 (d, 1H, J = 5.2 Hz), 8.24 (s, 1H), 8.06 (br s, 1H), 7.45 (d, 1H, J = 5.2 Hz), 7.25 (s, 4H), 6.64 (d, 1H, J = 11.6 Hz), 5.74 (dt, 1H, $J_1 = 11.6$ Hz, $J_2 = 6.7$ Hz), 4.42 (t, 2H, $J = 6.4$ Hz), 2.97– 2.90 (m, 1H), 1.28 (d, 6H, $J = 6.9$ Hz); ¹³C NMR (100 MHz, CDCl₃) δ 163.0, 151.3, 149.0, 148.2, 145.9, 133.8, 132.0, 128.8, 126.7, 126.5, 126.3, 122.9, 38.0, 33.9, 23.9; HRMS (ESI) m/z [M + H]⁺ calcd for $C_{18}H_{20}ClN_2O$ 315.1264, found 315.1252.

(Z)-4-Chloro-N-(3-(p-tolyl)allyl)picolinamide (4d). Compound 4d ((Z) major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 56% (40 mg, $E/Z =$ 17:83); IR (DCM) 2926, 1677, 1525, 1260 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.45 (d, 1H, J = 5.2 Hz), 8.23 (d, 1H, J = 2.0 Hz), 8.07 (br s, 1H), 7.45 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 7.20 (s, 4H), 6.63 (d, 1H, J = 11.5 Hz), 5.73 (dt, 1H, J₁ = 11.5 Hz, J₂ = 6.7 Hz), 4.41 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.8$ Hz), 2.38 (s, 3H); ¹³C NMR (100 MHz, CDCl3) δ 163.0, 151.3, 149.0, 145.9, 137.2, 133.4, 132.0, 129.1, 128.7,

126.8, 126.4, 122.9, 38.0, 21.3; HRMS (ESI) m/z [M + Na]⁺ calcd for $C_{16}H_{15}CIN_2NaO$ 309.0771, found 309.0772.

(Z)-4-Chloro-N-(3-(4-ethylphenyl)allyl)picolinamide (4e). Compound 4e ((Z) major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 50% (38 mg, $E/Z = 16:84$); IR (DCM) 2964, 1675, 1525 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.45 (d, 1H, J = 5.2 Hz), 8.24 (d, 1H, J = 2.0 Hz), 8.07 (br s, 1H), 7.45 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 7.25 (s, 4H), 6.65 (d, 1H, $J = 11.5$ Hz), 5.74 (dt, 1H, $J_1 = 11.5$ Hz, $J_2 = 6.7$ Hz), 4.41(td, 2H, J_1 = 6.7 Hz, J_2 = 1.8 Hz), 2.67 (q, 2H, J = 7.6 Hz), 1.26 (t, 3H, J = 7.6 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 163.0, 151.3, 149.0, 145.9, 143.6, 133.6, 132.0, 128.8, 127.9, 126.8, 126.4, 122.9, 38.0, 28.6, 15.6; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₇H₁₈ClN₂O 301.1108, found 301.1100.

(Z)-4-Chloro-N-(3-(4-pentylphenyl)allyl)picolinamide (4f). Compound 4f $((Z)$ major isomer) was obtained after purification by column chromatography on neutral alumina (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 57% (49 mg, $E/Z = 20:80$); IR (DCM) 2928, 1679, 1523 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.45 (d, 1H, J = 5.2 Hz), 8.24 (d, 1H, J = 2.0 Hz), 8.07 (br s, 1H), 7.45 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 7.23 $(d, 2H, J = 8.5 Hz)$, 7.20 $(d, 2H, J = 8.5 Hz)$, 6.64 $(d, 1H, J = 11.6)$ Hz), 5.74 (dt, 1H, $J = 11.6$ Hz, $J = 6.7$ Hz), 4.41 (td, 2H, $J = 1.8$ Hz, J $= 6.7$ Hz), 2.62 (t, 2H, $J = 7.6$ Hz), 1.67–1.60 (m, 2H), 1.39–1.32 (m, 4H), 0.92 (t, 3H, J = 6.9 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 163.0, 151.3, 149.0, 145.9, 142.3, 137.2, 133.6, 132.0, 130.6, 128.7, 128.5, 126.7, 126.3, 122.9, 38.0, 35.7, 31.5, 31.1, 22.6, 14.1; HRMS (ESI) m/z $[M + H]^{+}$ calcd for $C_{20}H_{24}C/N$, O 343.1577, found 343.1562.

(Z)-4-Chloro-N-(3-(4-hexylphenyl)allyl)picolinamide (4g). Compound $4g$ $((Z)$ major isomer) was obtained after purification by column chromatography on neutral alumina (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 50% (45 mg, E/Z = 20:80); IR (DCM) 2929, 1674, 1508, 1458 cm⁻¹;
¹H NMR (400 MHz, CDCL) δ 8 45 (d, 1H, I – 5 2 Hz), 8 24 (d, 1H, I ¹H NMR (400 MHz, CDCl₃) δ 8.45 (d, 1H, J = 5.2 Hz), 8.24 (d, 1H, J $= 1.9$ Hz), 8.07 (br s, 1H), 7.44 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 1.9$ Hz), 7.22 $(d, 2H, J = 8.4 Hz)$, 7.20 $(d, 1H, J = 8.4 Hz)$, 6.63 $(d, 1H, J = 11.5$ Hz), 5.73 (dt, 1H, $J_1 = 11.5$ Hz, $J_2 = 6.7$ Hz), 4.41 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.5$ Hz), 2.62 (t, 2H, J = 7.6 Hz), 1.65−1.59 (m, 2H), 1.38−1.28 (m, 6H), 0.91 (t, 3H, J = 6.4 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 163.0, 151.3, 149.0, 145.9, 142.3, 133.6, 132.0, 128.7, 128.4, 126.7, 126.3, 122.9, 38.0, 35.7, 31.7, 31.4, 29.0, 22.6, 14.1; HRMS (ESI) m/z $[M + H]^{+}$ calcd for $C_{21}H_{26}C/N_{2}O$ 357.1734, found 357.1718.

(Z)-4-Chloro-N-(3-(3,4-dimethylphenyl)allyl)picolinamide (4h). Compound $4h$ $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 52% (39 mg, E/Z = 12:88); IR (DCM) 1717, 1522, 1281, 1106 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.45 (d, 1H, J = 5.2 Hz), 8.24 (d, 1H, J = 2.0 Hz), 8.07 (br s, 1H), 7.44 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 7.15 (d, 1H, J = 7.6 Hz), 7.07−7.04 (m, 2H), 6.61 (d, 1H, J = 11.6 Hz), 5.72 (dt, 1H, $J_1 = 11.6$ Hz, $J_2 = 6.6$ Hz), 4.41 (td, 2H, $J_1 = 6.6$ Hz, $J_2 = 1.4$ Hz), 2.29 (s, 3H), 2.29 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 163.0, 151.3, 149.0, 145.9, 136.6, 135.9, 133.9, 132.0, 130.1, 129.6, 126.6, 126.3, 126.2, 122.9, 38.0, 19.9, 19.6; HRMS (ESI) m/z [M + H ⁺ calcd for C₁₇H₁₈ClN₂O 301.1108, found 301.1100.

(Z)-4-Chloro-N-(3-(2,3-dihydrobenzo[b][1,4]dioxin-6-yl)allyl) picolinamide (4i). Compound 4i $((Z)$ major isomer) was obtained after purification by column chromatography on neutral alumina (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/ hexanes = 1:4); yield 60% (50 mg, E/Z = 28:72); IR (DCM) 3377, 1674, 1580, 1293 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.45 (d, 1H, J $= 5.3$ Hz), 8.23 (d, 1H, J = 2.1 Hz), 8.07 (br s, 1H), 7.45 (dd, 1H, J₁ = 5.2 Hz, $J_2 = 2.1$ Hz), 6.87 (d, 1H, J = 8.2 Hz), 6.83–6.77 (m, 2H), 6.53 (d, 1H, J = 11.5 Hz), 5.67 (dt, 1H, J₁ = 11.5 Hz, J₂ = 6.7 Hz), 4.39 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.8$ Hz), 4.28 (s, $4H$); ¹³C NMR (100 MHz, CDCl3) δ 163.0, 151.3, 149.0, 145.9, 143.2, 143.0, 131.4, 129.8, 126.4, 126.3, 122.9, 122.2, 117.6, 117.2, 64.4, 64.3, 38.0; HRMS (ESI) m/z $[M + H]^+$ calcd for $C_{17}H_{16}C/N_2O_3$ 331.0849, found 331.0839.

(Z)-4-Chloro-N-(3-(3,5-dimethylphenyl)allyl)picolinamide (4j). Compound $4i$ $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.51$ (EtOAc/hexanes = 1:4); yield 53% (40 mg, $E/Z = 28:72$); IR (DCM) 3391, 1675, 1523, 1289 cm⁻¹; ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3)$ δ 8.45 (d, 1H, J = 5.2 Hz), 8.24 (d, 1H, J = 1.8 Hz), 8.06 (br s, 1H), 7.44 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 6.94– 6.92 (m, 3H), 6.61 (d, 1H, J = 11.6 Hz), 5.73 (dt, 1H, J_1 = 11.6 Hz, J_2 = 6.6 Hz), 4.41 (td, 2H, J_1 = 6.6 Hz, J_2 = 1.8 Hz), 2.35 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 163.0, 151.3, 149.0, 145.9, 137.9, 136.2, 132.2, 129.0, 127.2, 126.6, 126.3, 122.9, 38.0, 21.4; HRMS (ESI) m/z $[M + Na]⁺$ calcd for $C_{17}H_{17}CIN_2NaO$ 323.0927, found 323.0923.

(Z)-Ethyl 3-(3-(4-Chloropicolinamido)prop-1-en-1-yl)benzoate (4k). Compound 4k $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes $= 30:70$) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 58% (50 mg, $E/Z = 23:77$); IR (DCM) 3319, 1718, 1531, 1487 cm⁻¹;
¹H NMR (400 MHz, CDCL) δ 8.45 (d, 1H J = 5.2 Hz), 8.23 (d, 1H J ¹H NMR (400 MHz, CDCl₃) δ 8.45 (d, 1H, J = 5.2 Hz), 8.23 (d, 1H, J $= 2.0$ Hz), 8.10 (br s, 1H), 7.99–7.97 (m, 2H), 7.51–7.42 (m, 3H), 6.69 (d, 1H, J = 11.5 Hz), 5.86 (dt, 1H, J₁ = 11.5 Hz, J₂ = 6.7 Hz), 4.43−4.37 (m, 4H), 1.42 (t, 3H, $J = 7.2$ Hz); ¹³C NMR (100 MHz, CDCl3) δ 166.4, 163.1, 151.2, 149.0, 145.9, 136.5, 132.9, 131.1, 130.7, 129.8, 128.8, 128.5, 128.4, 126.4, 122.9, 61.1, 37.8, 14.3; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₈H₁₈ClN₂O₃ 345.1006, found 345.0994.

(Z)-4-Chloro-N-(3-(thiophene-2-yl)allyl)picolinamide (4l). Compound 41 $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 63% (44 mg, $E/Z = 37:63$); IR (DCM) 3369, 1674, 1526, 1287 cm⁻¹; ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3)$ δ 8.46 (d, 1H, J = 5.0 Hz), 8.24 (d, 1H, J = 1.8 Hz), 8.17 (br s, 1H), 7.45 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 7.36– 7.34 (m, 1H), 7.08–7.06 (m, 2H), 6.71 (d, 1H, $J = 11.5$ Hz), 5.69 (dt, 1H, $J_1 = 11.5$ Hz, $J_2 = 6.7$ Hz), 4.49 (td, 2H, $J_1 = 6.7$ Hz, $J_2 = 1.8$ Hz); 13 C NMR (100 MHz, CDCl₃) δ 163.1, 151.2, 149.0, 145.9, 139.2, 128.2, 127.2, 126.4, 126.2, 125.7, 124.2, 122.9, 38.3; HRMS (ESI) m/z $[M + H]^{+}$ calcd for $C_{13}H_{12}CIN_{2}OS$ 279.0359, found 279.0348.

(Z)-N-(3-(5-Bromopyridin-2-yl)allyl)-4-chloropicolinamide (4m). Compound $4m$ $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a pale yellow liquid: $R_f = 0.45$ (EtOAc/hexanes = 1:4); yield 54% (48 mg, E/Z = 23:77); IR (DCM) 3386, 1663, 1524, 1217 cm⁻¹; ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3)$ δ 8.74 (d, 1H, J = 2.2 Hz), 8.69 (br s, 1H), 8.47 (d, 1H, $J = 5.2$ Hz), 8.23 (dd, 1H, $J_1 = 2.0$ Hz, $J_2 = 0.4$ Hz), 7.81 (dd, 1H, $J_1 = 8.3$ Hz, $J_2 = 2.4$ Hz), 7.44 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.0$ Hz), 7.15 (d, 1H, J = 8.3 Hz), 6.52 (d, 1H, J = 11.6 Hz), 6.10 (dt, 1H, J₁ = 11.6 Hz, L = 6.8 Hz, L = 1.6 Hz): ¹³C 11.6 Hz, $J_2 = 6.8$ Hz), 4.64 (td, 2H, $J_1 = 6.8$ Hz, $J_2 = 1.6$ Hz); NMR (100 MHz, CDCl₃) δ 163.1, 154.0, 151.7, 150.4, 149.1, 145.8, 139.0, 133.3, 129.3, 126.2, 125.6, 123.0, 118.8, 37.9; HRMS (ESI) m/z $[M + H]^{+}$ calcd for $C_{14}H_{12}BrClN_3O$ 351.9852, found 351.9844.

(Z)-4-Chloro-N-(3-(6-fluoropyridin-3-yl)allyl)picolinamide (4n). Compound $4n$ $((Z)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.45$ (EtOAc/hexanes = 1:4); yield 50% (37) mg, E/Z = 18:82); IR (DCM) 3381, 1717, 1678, 1522 cm⁻¹; ¹H NMR (400 MHz, CDCl3) δ 8.47−8.45 (m, 1H), 8.22 (br s, 1H), 8.16 (br s, 1H), 8.13 (br s, 1H), 7.78 (t, 1H, J = 8.0 Hz), 7.47−7.46 (m, 1H), 6.98 (dd, 1H, $J_1 = 8.0$ Hz, $J_2 = 2.8$ Hz), 6.58 (d, 1H, $J = 11.6$ Hz), 5.95−5.90 (m, 1H), 4.35−4.32 (m, 2H); 13C NMR (100 MHz, CDCl₃) δ 163.2, 162.6 (d, J_{C−F} = 238.6 Hz), 151.0, 149.0, 147.6 (d, J_{C-F} = 14.6 Hz), 146.0, 141.1 (d, J_{C-F} = 7.8 Hz), 130.1, 130.0 (d, J_{C-F} $=$ 4.7 Hz), 127.0, 126.5, 123.0, 109.3 (d, J_{C-F} = 37.2 Hz), 37.7; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₄H₁₂ClFN₃O 292.0653, found 292.0644.

(Z)-N-(2-(4-Methylbenzyl)-3-(p-tolyl)allyl)picolinamide (5a). Compound 5a was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.50 (EtOAc/hexanes = 1:4); yield 43% (39 mg); IR (DCM) 3442, 1675, 1521, 1433 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.54–8.52 (m, 1H), 8.21 (dt, 1H, $J_1 = 7.8$ Hz, $J_2 = 1.0$ Hz), 7.99 (br s, 1H), 7.86 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.46–7.42 (m, 1H), 7.21–7.19 (m, 4H), 7.16 (d, 2H, J = 7.8 Hz), 7.11 (d, 2H, J = 7.8 Hz), 6.56 (s, 1H), 4.26 $(d, 2H, J = 5.8 \text{ Hz})$, 3.55 (s, 2H), 2.36 (s, 3H), 2.32 (s, 3H); ¹³C NMR (100 MHz, CDCl3) δ 164.3, 149.8, 148.0, 137.4, 137.3, 136.6, 136.1, 135.8, 134.0, 130.4, 129.2, 129.0, 129.0, 128.7, 126.1, 122.2, 42.1, 39.3, 21.2, 21.1; HRMS (ESI) m/z $[M + H]^+$ calcd for $C_{24}H_{25}N_2O$ 357.1967, found 357.1982.

(Z)-N-(2-(4-Ethylbenzyl)-3-(4-ethylphenyl)allyl)picolinamide (5b). Compound 5b was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.50 (EtOAc/hexane = 1:4); yield 36% (29 mg); IR (DCM) 3389, 1679, 1512, 1463 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.54–8.52 (m, 1H), 8.21 (dt, 1H, J_1 = 7.8 Hz, J_2 = 1.0 Hz), 7.99 (br s, 1H), 7.86 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.45−7.42 (m, 1H), 7.25−7.18 (m, 6H), 7.14 (d, 2H, $J = 8.1$ Hz), 6.59 (s, 1H), 4.27 (d, 2H, $J = 5.8$ Hz), 3.56 (s, 2H), 2.69−2.60 (m, 4H), 1.27−1.21 (m, 6H); 13C NMR (100 MHz, CDCl₃) δ 164.3, 149.8, 148.0, 143.0, 142.2, 137.4, 137.3, 136.3, 134.3, 130.5, 129.0, 128.8, 128.0, 127.9, 126.1, 122.2, 42.2, 39.3, 28.6, 28.5, 15.6, 15.6; HRMS (ESI) m/z [M + H]⁺ calcd for C₂₆H₂₉N₂O 385.2280, found 385.2290.

(Z)-N-(2-(4-Chlorobenzyl)-3-(4-chlorophenyl)allyl)picolinamide (5c). Compound 5c was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 40% (40 mg); IR (DCM) 3443, 1637, 1489 cm[−]¹ ; 1 H NMR (400 MHz, CDCl3) δ 8.54−8.52 (m, 1H), 8.18 (dt, 1H, J₁ = 7.8 Hz, J₂ = 1.0 Hz), 7.96 (br s, 1H), 7.87 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.46−7.44 (m, 1H), 7.33 $(d, 2H, J = 8.5 Hz),$ 7.27–7.21 (m, 6H), 6.50 (s, 1H), 4.23 (d, 2H, J = 6.0 Hz), 3.55 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 164.3, 149.5, 148.1, 138.4, 137.4, 137.4, 135.1, 132.9, 132.3, 130.4, 130.1, 129.6, 128.7, 128.6, 126.3, 122.2, 41.8, 39.2; HRMS (ESI) m/z [M + H]⁻ calcd for $C_{22}H_{19}Cl_2N_2O$: 397.0874, found 397.0865.

(Z)-Dimethyl 4,4′-(2-(Picolinamidomethyl)prop-1-ene-1,3-diyl) dibenzoate (5d). Compound 5d was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 50% (56 mg); IR (DCM) 3377, 1718, 1678, 1522 cm[−]¹ ; 1 H NMR (400 MHz, CDCl₃) δ 8.51–8.49 (m, 1H), 8.17 (dt, 1H, J_1 = 7.8 Hz, J_2 = 1.0 Hz), 8.03 (d, 2H, J = 8.3 Hz), 7.99 (br s, 1H), 7.97 (d, 2H, J = 8.3 Hz), 7.86 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.45–7.38 (m, 1H), 7.37 (d, 4H, J = 8.2 Hz), 6.56 (s, 1H), 4.27 (d, 2H, J = 5.9 Hz), 3.92 (s, 3H), 3.91 (s, 3H), 3.65 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 167.0, 166.8, 164.3, 149.4, 148.0, 144.3, 141.4, 139.5, 137.4, 130.1, 129.9, 129.7, 129.1, 128.8, 128.6, 128.4, 126.3, 122.2, 52.1, 52.1, 42.3, 39.3; HRMS (ESI) m/z [M + H]⁺ calcd for C₂₆H₂₅N₂O₅ 445.1763, found 445.1748.

(Z)-N-(2-(4-Isopropylbenzyl)-3-(4-isopropylphenyl)allyl) picolinamide (5e). Compound 5e was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 37% (39 mg), IR (DCM) 3390, 1680, 1518, 1433 cm[−]¹ ; 1 H NMR (400 MHz, CDCl₃) δ 8.54–8.52 (m, 1H), 8.21 (dt, 1H, J₁ = 7.8 Hz, J₂ = 1.0 Hz), 7.99 (br s, 1H), 7.86 (td, 1H, J₁ = 7.7 Hz, J₂ = 1.7 Hz), 7.45−7.42 (m, 1H), 7.28−7.19 (m, 6H), 7.16 (d, 2H, J = 8.1 Hz), 6.59 (s, 1H), 4.28 $(d, 2H, J = 5.7 Hz)$, 3.56 (s, 1H), 2.98–2.85 (m, 2H), 1.26 (d, 6H, J = 6.9 Hz), 1.24 (d, 6H, J = 6.9 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 164.3, 149.8, 148.0, 147.6, 146.8, 137.3, 137.3, 136.4, 134.4, 130.6, 128.9, 128.7, 126.5, 126.4, 126.1, 122.2, 42.3, 39.3, 33.8, 33.7, 24.1, 24.0; HRMS (ESI) m/z [M + H]⁺ calcd for C₂₈H₃₃N₂O 413.2593, found 413.2594.

(Z)-N-(2-(3,4-Dimethylbenzyl)-3-(3,4-dimethylphenyl)allyl) picolinamide (5f). Compound 5f was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a yellow liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 44% (43 mg); IR (DCM) 3396, 1678, 1520, 1433 cm[−]¹ ; 1 H NMR (400 MHz, CDCl₃) δ 8.53–8.51 (m, 1H), 8.20 (dt, 1H, J_1 = 7.8 Hz, J_2 = 1.0 Hz), 7.97 (br s, 1H), 7.86 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.45–7.42 (m, 1H), 7.13−7.03 (m, 6H), 6.55 (s, 1H), 4.28 (d, 2H, J = 5.7 Hz), 3.52 (s, 2H), 2.27 (s, 6H), 2.22 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 164.2, 149.9, 148.0, 137.2, 136.6, 136.6, 136.4, 135.3, 134.5, 134.4, 130.4, 130.4, 130.1, 129.7, 129.6, 126.4, 126.1, 126.0, 122.1, 42.3, 39.4,

19.8, 19.8, 19.5, 19.4; HRMS (ESI) m/z [M + H]⁺ calcd for $C_{26}H_{29}N_{2}O$ 385.2280, found 385.2292.

(Z)-N-(3-(2,3-Dihydrobenzo[b][1,4]dioxin-6-yl)-2-((2,3 dihydrobenzo[b][1,4]dioxin-6-yl)methyl)allyl)picolinamide (5g). Compound 5g was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.50 (EtOAc/hexanes = 1:4); yield 46% (52 mg); IR (DCM) 3441, 1637, 1505, 1284, 749 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.55– 8.53 (m, 1H), 8.20 (dt, 1H, J_1 = 7.8 Hz, J_2 = 1.0 Hz), 7.98 (br s, 1H), 7.85 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.45−7.42 (m, 1H), 6.85−6.83 (m, 2H), 6.81–6.78 (m, 3H), 6.75 (dd, 1H, $J_1 = 8.3$ Hz, $J_2 = 1.8$ Hz), 6.47 (s, 1H), 4.30−4.22 (m, 10H), 3.45 (s, 2H); 13C NMR (100 MHz, CDCl3) δ 164.2, 149.8, 148.0, 143.4, 143.2, 142.6, 142.1, 137.3, 136.9, 132.4, 130.4, 129.9, 126.1, 122.2, 122.1, 122.0, 117.6, 117.6, 117.2, 117.1, 64.5, 64.4, 64.3, 64.3, 41.8, 39.2; HRMS (ESI) m/z $[M + H]$ ⁺ calcd for $C_{26}H_{25}N_2O_5$ 445.1763, found 445.1775.

(Z)-N-(2-(3,4-Dichlorobenzyl)-3-(3,4-dichlorophenyl)allyl) picolinamide (5h). Compound 5h was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 45% (53 mg); IR (DCM) 3384, 1738, 1676, 1468 cm[−]¹ ; 1 H NMR (400 MHz, CDCl₃) δ 8.53–8.51 (m, 1H), 8.17 (dt, 1H, J₁ = 7.8 Hz, J₂ = 1.0 Hz), 7.96 (br s, 1H), 7.87 (td, 1H, J₁ = 7.7 Hz, J₂ = 1.7 Hz), 7.47–7.45 (m, 1H), 7.44 (d, 1H, $J = 8.2$ Hz), 7.40 (d, 1H, $J = 1.7$ Hz), 7.35 (d, 2H, J $= 8.3$ Hz), 7.17 (dd, 1H, $J_1 = 8.3$ Hz, $J_2 = 2.0$ Hz), 7.12 (dd, 1H, $J_1 =$ 8.1 Hz, $J_2 = 2.0$ Hz), 6.45 (s, 1H), 4.24 (dd, 2H, $J_1 = 6.1$ Hz, $J_2 = 0.7$ Hz), 3.53 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 164.3, 149.3, 148.1, 139.2, 139.0, 137.4, 136.5, 132.6, 132.5, 131.2, 130.9, 130.7, 130.6, 130.5, 130.4, 128.7, 128.4, 128.1, 126.4, 122.2, 41.5, 39.2; HRMS (ESI) m/z [M + H]⁺ calcd for C₂₂H₁₇Cl₄N₂O 465.0095, found 465.0081.

(Z)-N-(2-(3,5-Dimethylbenzyl)-3-(3,5-dimethylphenyl)allyl) picolinamide (5i). Compound 5i was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 47% (46 mg); IR (DCM) 3388, 1679, 1520, 1464 cm[−]¹ ; 1 H NMR (400 MHz, CDCl₃) δ 8.54–8.52 (m, 1H), 8.21 (dt, 1H, J_1 = 7.8 Hz, J_2 = 1.0 Hz), 7.97 (br s, 1H), 7.86 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.45–7.42 (m, 1H), 6.94−6.91 (m, 5H), 6.83 (s, 1H), 6.54 (s, 1H), 4.30 (d, 2H, J = 5.8 Hz), 3.51 (s, 2H), 2.33 (s, 6H), 2.28 (s, 6H); 13C NMR (100 MHz, CDCl₃) δ 164.2, 149.9, 147.9, 139.1, 137.9, 137.8, 137.6, 137.2, 136.9, 130.6, 128.5, 127.9, 126.9, 126.6, 126.0, 122.1, 42.5, 39.5, 21.3, 21.3; HRMS (ESI) m/z [M + H]⁺ calcd for C₂₆H₂₉N₂O 385.2280, found 385.2289.

(Z)-Diethyl 3,3′-(2-(picolinamidomethyl)prop-1-ene-1,3-diyl) dibenzoate (5j). Compound 5j was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 41% (49 mg); IR (DCM) 1716, 1680, 1521, 1204 cm[−]¹ ; 1 H NMR (400 MHz, CDCl₃) δ 8.52–8.50 (m, 1H), 8.18 (dt, 1H, J₁ = 7.8 Hz, J₂ = 1.0 Hz), 8.02 (br s, 1H), 7.97−7.91 (m, 3H), 7.89−7.83 (m, 2H), 7.51 (d, 2H, J = 7.7 Hz), 7.45−7.37 (m, 3H), 6.56 (s, 1H), 4.39 (q, 2H, J = 7.1 Hz), 4.37 (q, 2H, J = 7.1 Hz), 4.27 (d, 2H, J = 5.9 Hz), 3.66 (s, 2H), 1.41 (t, 3H, $J = 7.1$ Hz), 1.38 (t, 3H, $J = 7.1$ Hz); ¹³C NMR (100 MHz, CDCl₃) δ 166.6, 166.5, 164.3, 149.5, 148.0, 139.1, 138.9, 137.3, 137.0, 133.7, 133.0, 130.8, 130.6, 130.2, 130.0, 129.9, 128.6, 128.5, 128.1, 127.8, 126.2, 122.2, 61.1, 61.0, 41.9, 39.3, 14.4, 14.3; HRMS (ESI) m/z $[M + H]^{+}$ calcd for $C_{28}H_{29}N_{2}O_{5}$ 473.2076, found 473.2093.

(Z)-N-(3-(2-Chloropyridin-4-yl)-2-((2-chloropyridin-4-yl)methyl) allyl)picolinamide $(5k)$. Compound $5k$ was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.45$ (EtOAc/hexanes = 1:4); yield 34% (34) mg); IR (DCM) 3371, 1672, 1527, 1465, 1386 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.52–8.51 (m, 1H), 8.39 (d, 1H, J = 5.1 Hz), 8.27 (d, 1H, $J = 5.1$ Hz), 8.16 (dt, 1H, $J_1 = 7.8$ Hz, $J_2 = 1.0$ Hz), 8.03 (br s, 1H), 7.88 (td, 1H, J_1 = 7.7 Hz, J_2 = 1.7 Hz), 7.49–7.45 (m, 1H), 7.29 (br s, 1H), 7.23 (br s, 1H), 7.21 (dd, 1H, $J_1 = 5.1$ Hz, $J_2 = 0.9$ Hz), 7.14 (dd, 1H, $J_1 = 5.1$ Hz, $J_2 = 1.4$ Hz), 6.42 (s, 1H), 4.27 (d, 2H, $J =$ 5.6 Hz), 3.58 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 164.5, 152.0, 150.8, 149.8, 148.9, 148.2, 147.1, 141.3, 137.6, 127.8, 126.6, 124.6,

124.0, 122.9, 122.3, 122.2, 41.1, 39.3; HRMS (ESI) m/z [M + H]⁺ calcd for $C_{20}H_{17}Cl_2N_4O$ 399.0779, found 399.0765.

(Z)-N-(3-(2-Chloropyridin-4-yl)-2-methylallyl)picolinamide (6a). Compound 6a was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.48 (EtOAc/hexanes = 1:4); yield 30% (22 mg); IR (DCM) 2925, 1713, 1423, 1364 cm[−]¹ ; 1 H NMR (400 MHz, CDCl3) δ 8.57−8.55 (m, 1H), 8.35 (d, 1H, $J = 5.1$ Hz), 8.22 (d, 1H, $J = 7.8$ Hz), 8.16 (br s, 1H), 7.89 (td, 1H, J¹ = 7.7 Hz, J² = 1.7 Hz), 7.49−7.45 (m, 1H), 7.25 (br s, 1H), 7.18 (dd, 1H, $J_1 = 5.1$ Hz, $J_2 = 0.9$ Hz), 6.38 (s, 1H), 4.30 (d, 2H, J = 6.1 Hz), 2.01 (d, 3H, J = 1.4 Hz); ¹³C NMR (100 MHz, CDCl3) δ 164.5, 151.8, 149.6, 149.4, 148.1, 148.0, 140.6, 137.5, 126.4, 124.9, 123.9, 122.3, 40.6, 22.4; HRMS (ESI) m/z [M + H]+ calcd for $C_{15}H_{15}CIN_3O$ 288.0904, found 288.0895.

(Z)-Diethyl 3,3′-(2-((4-chloropicolinamido)methyl)prop-1-ene-1,3-diyl)dibenzoate (7a). Compound 7a was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.50 (EtOAc/hexanes = 1:4); yield 40% (51 mg); IR (DCM) 1716, 1679, 1521, 1282 cm⁻¹; ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3)$ δ 8.39 (d, 1H, J = 5.2 Hz), 8.17 (d, 1H, J = 1.9 Hz), 7.95−7.94 (m, 3H), 7.91−7.89 (m, 2H), 7.50 (dd, 2H, J¹ = 7.7 Hz, $J_2 = 1.3$ Hz), 7.46–7.36 (m, 3H), 6.58 (s, 1H), 4.39 (q, 4H, $J = 7.1$ Hz), 4.27 (d, 2H, $J = 6.0$ Hz), 3.65 (s, 2H), 1.40 (t, 6H, $J = 7.1$ Hz); 13 C NMR (100 MHz, CDCl₃) δ 166.6, 166.4, 163.1, 151.0, 148.9, 145.8, 139.1, 138.5, 136.9, 133.5, 133.0, 130.8, 130.6, 130.2, 130.1, 129.8, 128.6, 128.5, 128.2, 127.8, 126.3, 122.8, 61.1, 61.0, 42.0, 39.4, 14.4, 14.3; HRMS (ESI) m/z [M + H]⁺ calcd for C₂₈H₂₈ClN₂O₅ 507.1687, found 507.1669.

(Z)-N-(3-(5-Bromopyridin-2-yl)-2-methylallyl)picolinamide (6b). Compound 6b was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.43 (EtOAc/hexanes = 1:4); yield 41% (34 mg); IR (DCM) 2929, 1716, 1674, 1520 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.91 (br s, 1H), 8.72 (d, 1H, $J = 2.3$ Hz), 8.56 (d, 1H, $J = 4.8$ Hz), 8.22 (d, 1H, J $= 7.8$ Hz), 7.85 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.77 (dd, 1H, $J_1 =$ 8.4 Hz, $J_2 = 2.4$ Hz), 7.43–7.40 (m, 1H), 7.10 (d, 1H, $J = 8.4$ Hz), 6.39 $(s, 1H)$, 4.49 (d, 2H, J = 6.6 Hz), 2.08 (d, 3H, J = 1.4 Hz); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3)$ δ 164.3, 154.5, 150.3, 150.2, 148.2, 142.5, 138.9, 137.2, 126.1, 126.0, 125.2, 122.3, 118.0, 40.6, 24.8; HRMS (ESI) m/z $[M + H]^+$ calcd for $C_{15}H_{15}BrN_3O$ 332.0398, found 332.0388.

(Z)-N-(3-(6-Fluoropyridin-3-yl)-2-methylallyl)picolinamide (6c). Compound 6c was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.43 (EtOAc/hexanes = 1:4); yield 41% (28 mg); IR (DCM) 2927, 1672, 1526, 1483 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.57–8.55 (m, 1H), 8.22 (d, 1H, $J = 7.8$ Hz), 8.12 (d, 1H, $J = 1.8$ Hz), 8.12 (br s, 1H), 7.89 (td, 1H, $J_1 = 7.7$ Hz, $J_2 = 1.7$ Hz), 7.78 (td, 1H, $J_1 = 8.1$ Hz, $J_2 = 2.4 \text{ Hz}$, 7.48–7.45 (m, 1H), 6.93 (dd, 1H, $J_1 = 8.4 \text{ Hz}$, $J_2 = 3.0 \text{ Hz}$), 6.43 (s, 1H), 4.24 (d, 2H, $J = 6.0 \text{ Hz}$), 2.01 (d, 3H, $J = 1.4 \text{ Hz}$); ¹³C NMR (100 MHz, CDCl₃) δ 164.5, 162.3 (d, J_{C−F} = 237.8 Hz), 149.5, 148.1, 147.4 (d, $J_{C-F} = 14.5$ Hz), 141.2 (d, $J_{C-F} = 7.8$ Hz), 137.8, 137.5, 130.8 (d, J_{C-F} = 4.6 Hz), 126.4, 123.7, 122.3, 109.1 (d, J_{C-F} = 37.1 Hz), 40.5, 22.2; HRMS (ESI) m/z [M + H]⁺ calcd for $C_{15}H_{15}FN_{3}O$ 272.1199, found 272.1188.

(Z)-N-(3-(5-Bromopyridin-2-yl)-2-methylallyl)-4-chloropicolinamide (6d). Compound 6d was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.40$ (EtOAc/hexanes = 1:4); yield 40% (37 mg); IR (DCM) 3384, 1675, 1515, 1464 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 8.95 (br s, 1H), 8.72 (d, 1H, J = 2.3 Hz), 8.46 (d, 1H, J = 5.2 Hz), 8.23 (d, 1H, J = 2.0 Hz), 7.78 (dd, 1H, J₁ = 8.4 Hz, J₂ = 2.4 Hz), 7.43 (dd, 1H, $J_1 = 5.2$ Hz, $J_2 = 2.1$ Hz), 7.09 (d, 1H, $J = 8.2$ Hz), 6.39 (s, 1H), 4.48 (d, 2H, $J = 6.7$ Hz), 2.08 (d, 3H, $J = 1.4$ Hz); ¹³C NMR (100 MHz, CDCl₃) δ 163.1, 154.4, 151.8, 150.2, 149.1, 145.7, 142.3, 139.0, 126.2, 126.1, 125.2, 123.0, 118.1, 40.7, 24.9; HRMS (ESI) m/z [M + H ⁺ calcd for C₁₅H₁₄BrClN₃O 366.0009, found 366.0021.

 $N₂N₆$ -Bis((E)-3-(4-methoxyphenyl)allyl)pyridine-2,6-dicarboxamide (8a). Compound 8a $((E)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: R_f = 0.50 (EtOAc/hexanes = 1:4); yield

40% (46 mg, E/Z = 67:33); IR (DCM) 3441, 1643, 1524, 1241 cm⁻¹;
¹H NMR (400 MHz, CDCL) δ 8.41 (d. 2H I = 7.8 Hz), 8.05 (t. 1H i ¹H NMR (400 MHz, CDCl₃) δ 8.41 (d, 2H, J = 7.8 Hz), 8.05 (t, 1H, J $= 7.8$ Hz), 8.00 (t, 1H, $J = 6.1$ Hz), 7.26 (d, 4H, $J = 8.8$ Hz), 6.82 (d, 4H, J = 8.8 Hz), 6.52 (d, 2H, J = 15.8 Hz), 6.13 (dt, 2H, J₁ = 15.8 Hz, $J_2 = 6.4$ Hz), 4.27 (td, 4H, $J_1 = 6.3$ Hz, $J_2 = 1.2$ Hz), 3.81 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 163.4, 159.3, 148.8, 139.0, 132.0, 129.1, 127.5, 125.3, 122.8, 114.0, 55.3, 41.8; HRMS (ESI) m/z [M + Na]⁺ calcd for $C_{27}H_{27}N_3NaO_4$ 480.1899, found 480.1916.

(E)-N-(3-(4-Methoxyphenyl)allyl)pyrazine-2-carboxamide (9a). Compound $9a$ $((E)$ major isomer) was obtained after purification by column chromatography on silica gel (EtOAc/hexanes = 30:70) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexane = 1:4); yield 35% (24 mg, $E/Z = 62:38$); IR (DCM) 3447, 1637, 1511 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 9.46 (d, 1H, J = 1.5 Hz), 8.78 (d, 1H, J = 2.5 Hz), 8.55 (dd, 1H, $J_1 = 2.5$ Hz, $J_2 = 1.5$ Hz), 7.96 (br s, 1H), 7.33 (d, 2H, J $= 8.8$ Hz), 6.87 (d, 2H, J = 8.8 Hz), 6.59 (d, 1H, J = 15.8 Hz), 6.17 (dt, 1H, $J_1 = 15.8$ Hz, $J_2 = 6.4$ Hz), 4.28 (td, 2H, $J_1 = 6.4$ Hz, $J_2 = 1.4$ Hz), 3.83 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 162.8, 159.4, 147.3, 144.5, 142.6, 132.3, 129.2, 127.6, 122.5, 114.0, 55.3, 41.6; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₅H₁₅N₃NaO₂ 292.1062, found 292.1051.

(E)-5-Methyl-N-(3-(p-tolyl)allyl)isoxazole-3-carboxamide (10a). Compound $10a$ $((E)$ major isomer) was obtained after purification by column chromatography on neutral alumina (EtOAc/hexanes = 35:65) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexane = 1:4); yield 78% (50 mg, E/Z = 98:2); IR (DCM) 3294, 1660, 1558, 1304 cm⁻¹;
¹H NMR (400 MHz, CDCL) δ 7.28 (d, 2H J = 8.0 Hz) 7.14 (d, 2H J ¹H NMR (400 MHz, CDCl₃) δ 7.28 (d, 2H, J = 8.0 Hz), 7.14 (d, 2H, J $= 8.0$ Hz), 7.02 (br s, 1H), 6.58 (d, 1H, $I = 15.8$ Hz), 6.48 (d, 1H, $I =$ 0.7 Hz), 6.20 (dt, 1H, $J_1 = 15.8$ Hz, $J_2 = 6.4$ Hz), 4.22 (td, 2H, $J_1 = 6.4$ Hz, $J_2 = 1.4$ Hz), 2.49 (d, 3H, $J = 0.7$ Hz), 2.35 (s, 3H); ¹³C NMR (100 MHz, CDCl3) δ 171.2, 159.0, 158.7, 137.7, 133.6, 132.6, 129.3, 126.3, 123.5, 101.5, 41.5, 21.2, 12.4; HRMS (ESI) m/z [M + H]⁺ calcd for $C_{15}H_{17}N_2O_2$ 257.1290, found 257.1281.

(E)-N-(3-(4-Methoxyphenyl)allyl)-5-methylisoxazole-3-carboxamide (10b). Compound 10b $((E)$ major isomer) was obtained after purification by column chromatography on neutral alumina (EtOAc/ hexanes = 35:65) as colorless liquid: R_f = 0.50 (EtOAc/hexane = 1:4); yield 60% (41 mg, E/Z = 98:2); IR (DCM) 3290, 1659, 1553, 1455 cm⁻¹;¹H NMR (400 MHz, CDCl₃) δ 7.32 (d, 2H, J = 8.7 Hz), 6.96 $(br s, 1H)$, 6.87 (d, 2H, J = 8.7 Hz), 6.56 (d, 1H, J = 15.8 Hz), 6.48 (br s, 1H), 6.12 (dt, 1H, $J_1 = 15.8$ Hz, $J_2 = 6.4$ Hz), 4.21 (td, 2H, $J_1 = 6.2$ Hz, $J_2 = 1.4$ Hz), 3.83 (s, 3H) 2.50 (d, 3H, $J = 0.7$ Hz); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 159.4, 159.0, 158.7, 132.3, 129.1, 127.6, 122.2, 114.0, 101.5, 55.3, 41.5, 12.4; HRMS (ESI) m/z [M + Na]⁺ calcd for $C_{15}H_{16}N_2NaO_3$ 295.1059, found 295.1048.

(E)-N-(3-(4-Bromophenyl)allyl)-5-methylisoxazole-3-carboxamide (10c). Compound 10c $((E)$ major isomer) was obtained after purification by column chromatography on neutral alumina (EtOAc/ hexanes = 35:65) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 69% (56 mg, E/Z = 98:2); IR (DCM) 3288, 1656, 1552, 1453 cm⁻¹;¹H NMR (400 MHz, CDCl₃) δ 7.44 (d, 2H, J = 8.5 Hz), 7.24 (d, 2H, $J = 8.5$ Hz), 7.05 (br s, 1H), 6.54 (d, 1H, $J = 15.8$ Hz), 6.48 (d, 1H, J = 0.9 Hz), 6.25 (dt, 1H, J₁ = 15.8 Hz, J₂ = 6.2 Hz), 4.22 (td, 2H, J_1 = 6.1 Hz, J_2 = 1.5 Hz), 2.49 (d, 3H, J = 0.8 Hz); ¹³C NMR $(100 \text{ MHz}, \text{CDCl}_3)$ δ 171.3, 159.0, 158.6, 135.3, 131.7, 131.3, 128.0, 125.5, 121.6, 101.5, 41.2, 12.4; HRMS (ESI) m/z [M + H]⁺ calcd for $C_{14}H_{14}BrN_2O_2$ 321.0239, found 321.0227.

(E)-Ethyl 3-(3-(5-methylisoxazole-3-carboxamido)prop-1-en-1 yl)benzoate (10d). Compound 10d $((E)$ major isomer) was obtained after purification by column chromatography on neutral alumina (EtOAc/hexanes = 35:65) as a colorless liquid: R_f = 0.50 (EtOAc/ hexanes = 1:4); yield 75% (59 mg, E/Z = 98:2); IR (DCM) 3335, 1716, 1681, 1546 cm⁻¹;¹H NMR (400 MHz, CDCl₃) δ 8.05 (s, 1H), 7.93 (d, 1H, $J = 7.8$ Hz), 7.55 (d, 1H, $J = 7.8$ Hz), 7.40 (t, 1H, $J = 7.7$ Hz), 7.06 (br s, 1H), 6.64 (d, 1H, $J = 15.9$ Hz), 6.48 (d, 1H, $J = 0.7$ Hz), 6.34 (dt, 1H, $J_1 = 15.9$ Hz, $J_2 = 6.1$ Hz), 4.39 (q, 2H, $J = 7.1$ Hz), 4.25 (td, 2H, $J_1 = 6.1$ Hz, $J_2 = 1.4$ Hz), 2.50 (s, 3H), 1.41 (t, 3H, J = 7.1 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 171.3, 166.4, 159.1, 158.7, 136.7, 131.5, 130.9, 130.6, 128.8, 128.6, 127.5, 126.0, 101.5, 61.1, 41.2, 14.4,

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12.4; HRMS (ESI) m/z [M + Na]⁺ calcd for C₁₇H₁₈N₂NaO₄ 337.1164, found 337.1152.

(E)-N-(3-(2,3-Dihydrobenzo[b][1,4]dioxin-6-yl)allyl)-5-methylisoxazole-3-carboxamide (10e). Compound 10e $((E)$ major isomer) was obtained after purification by column chromatography on neutral alumina (EtOAc/hexanes = 35:65) as a colorless solid; $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 70% (53 mg, E/Z = 98:2); mp 115−117 °C; IR (DCM) 3434, 1673, 1508, 1308 cm⁻¹;¹H NMR (400 MHz, CDCl₃) δ 7.00 (br s, 1H), 6.90–6.85 (m 2H), 6.81 (d, 1H, J = 8.3 Hz), 6.48 (d, 1H, J = 15.8 Hz), 6.47 (s, 1H), 6.09 (dt, 1H, J₁ = 15.8 Hz, $J_2 = 6.4$ Hz), 4.26 (s, 4H), 4.19 (td, 2H, $J_1 = 6.2$ Hz, $J_2 = 1.4$ Hz), 2.49 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.2, 158.9, 158.7, 143.5, 143.4, 132.1, 130.2, 122.9, 119.9, 117.3, 115.0, 101.5, 64.4, 64.3, 41.4, 12.4; HRMS (ESI) m/z [M + H]⁺ calcd for C₁₆H₁₇N₂O₄ 301.1188, found 301.1176.

(E)-5-Methyl-N-(3-(3-nitrophenyl)allyl)isoxazole-3-carboxamide (10f). Compound 10f $((E)$ major isomer) was obtained after purification by column chromatography on neutral alumina (EtOAc/ hexanes = 35:65) as a colorless liquid: $R_f = 0.50$ (EtOAc/hexanes = 1:4); yield 64% (46 mg, E/Z = 98:2); IR (DCM) 3325, 1674, 1529, 1457 cm[−]¹ ; 1 H NMR (400 MHz, CDCl3) δ 8.22 (s, 1H), 8.10 (dd, 1H, $J_1 = 8.1$ Hz, $J_2 = 1.7$ Hz), 7.68 (d, 1H, $J = 7.7$ Hz), 7.50 (t, 1H, $J = 8.0$ Hz), 7.11 (br s, 1H), 6.66 (d, 1H, J = 15.9 Hz), 6.49 (s, 1H), 6.42 (dt, 1H, $J_1 = 15.9$ Hz, $J_2 = 5.9$ Hz), 4.29–4.27 (m, 2H), 2.51 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 171.4, 159.1, 158.6, 148.6, 138.2, 132.2, 129.9, 129.5, 128.3, 122.4, 121.1, 101.5, 41.0, 12.4; HRMS (ESI) m/z $[M + H]^{+}$ calcd for $C_{14}H_{14}N_{3}O_{4}$ 288.0984, found 288.0971.

■ ASSOCIATED CONTENT

6 Supporting Information

The Supporting Information is available free of charge on the [ACS Publications website](http://pubs.acs.org) at DOI: [10.1021/acs.joc.7b00535.](http://pubs.acs.org/doi/abs/10.1021/acs.joc.7b00535)

X-ray structure of compound 10e (Figures S1 and S2); brief X-ray structure data of compound 10e (Table S1); 1 H/¹³C NMR charts; crude NMR spectra of reactions revealing the observed E/Z ratios ([PDF\)](http://pubs.acs.org/doi/suppl/10.1021/acs.joc.7b00535/suppl_file/jo7b00535_si_001.pdf)

X-ray structure data of compound 10e [\(CIF\)](http://pubs.acs.org/doi/suppl/10.1021/acs.joc.7b00535/suppl_file/jo7b00535_si_002.cif)

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Notes

The authors declare no competing financial interest.

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